



***От организации
хроматина к
пониманию
функционирования
геномов эукариот***

Алексей Константинович Шайтан

д.ф.-м.н., профессор, чл.-корр. РАН

кафедра биоинженерии

биологический факультет

МГУ имени М.В.Ломоносова

***Лекция 6.
Экспериментальная эпигеномика.***

Апрель 2024

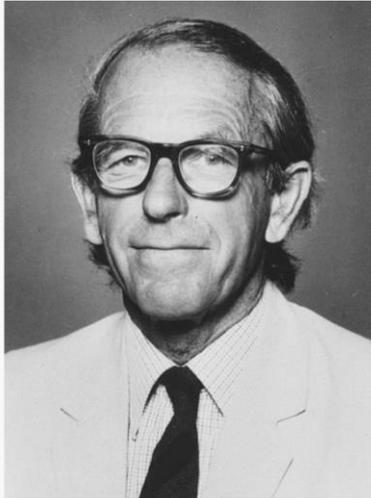
Содержание лекции

- Методы секвенирования 1,2,3 поколений.
- Проект геном человека.
- Методы секвенирования в эпигенетике: бисульфитное, Nano-pore, Chip-Seq, Chip-Exo, Mnase-seq, ATAC-seq, CUT&TAG, CUT&RUN, Ribo-seq, Hi-C,
- От проекта геном человека к 4Д-нуклеому (включая ENCODE)
- GWAS/EWAS

Секвенирование по Сенгеру (1-ое поколение)

Frederick Sanger

OM CH CBE FRS FAA



Born 13 August 1918
[Rendcomb](#), Gloucestershire,
 England

Died 19 November 2013 (aged 95)
[Cambridge](#), England^[1]

1958		Frederick Sanger (1918-2013)	 United Kingdom	"for his work on the structure of proteins , especially that of insulin"
------	--	--	--	--

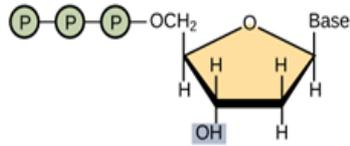
1980		Paul Berg (b. 1926)	 United States	"for his fundamental studies of the biochemistry of nucleic acids, with particular regard to recombinant-DNA "
		Walter Gilbert (b. 1932)	 United States	
		Frederick Sanger (1918-2013)	 United Kingdom	"for their contributions concerning the determination of base sequences in nucleic acids"

1951-52 – последовательность инсулина

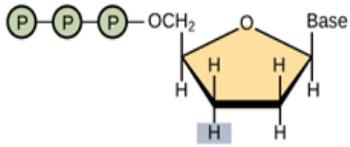
Метод Сенгера

1977 год

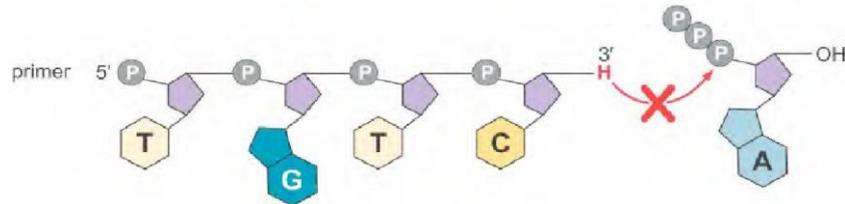
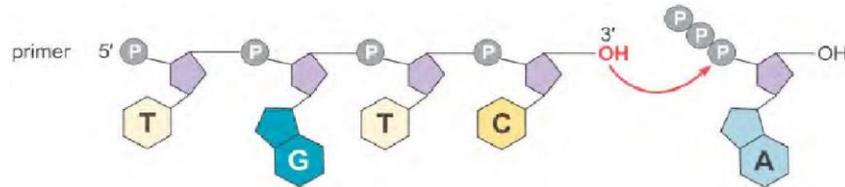
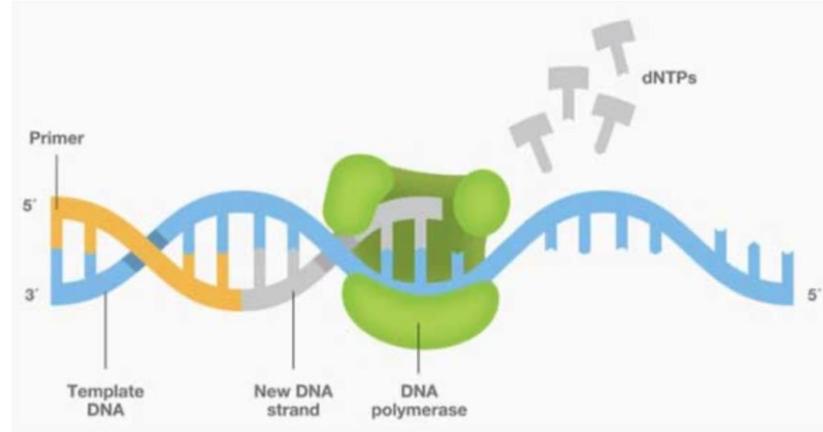
Chain termination method



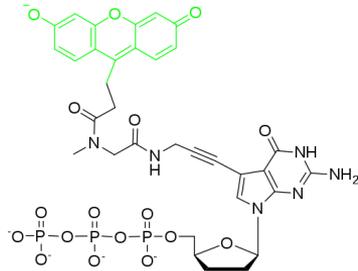
Deoxynucleotide (dNTP)



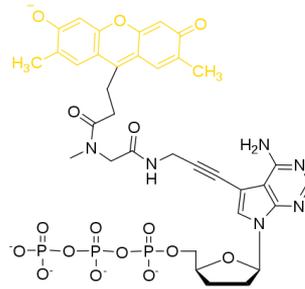
Dideoxynucleotide (ddNTP)



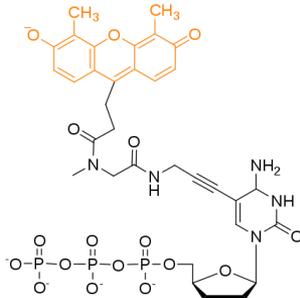
dye-terminating sequencing



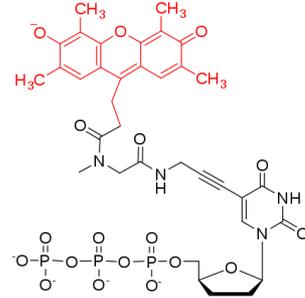
G-505



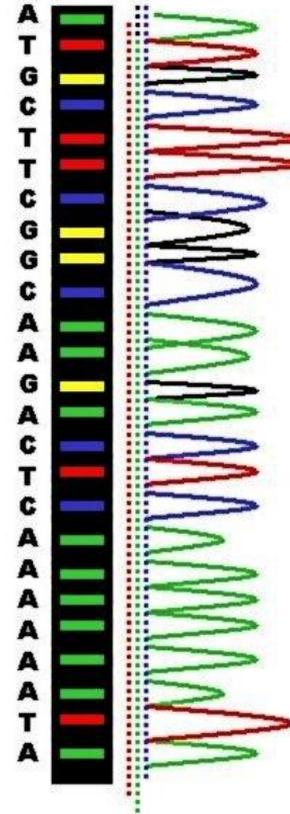
A-512



C-519



T-526

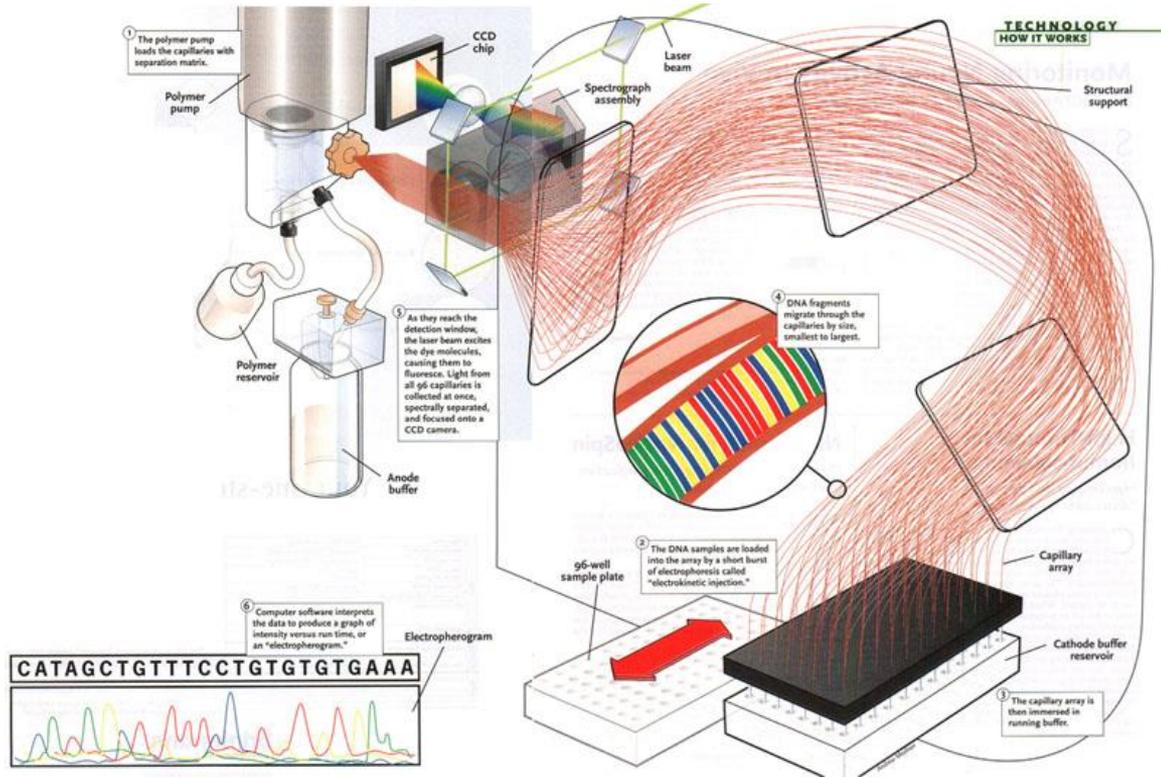


ддНТФ с флуоресцентными метками

хроматограмма

Первое поколение

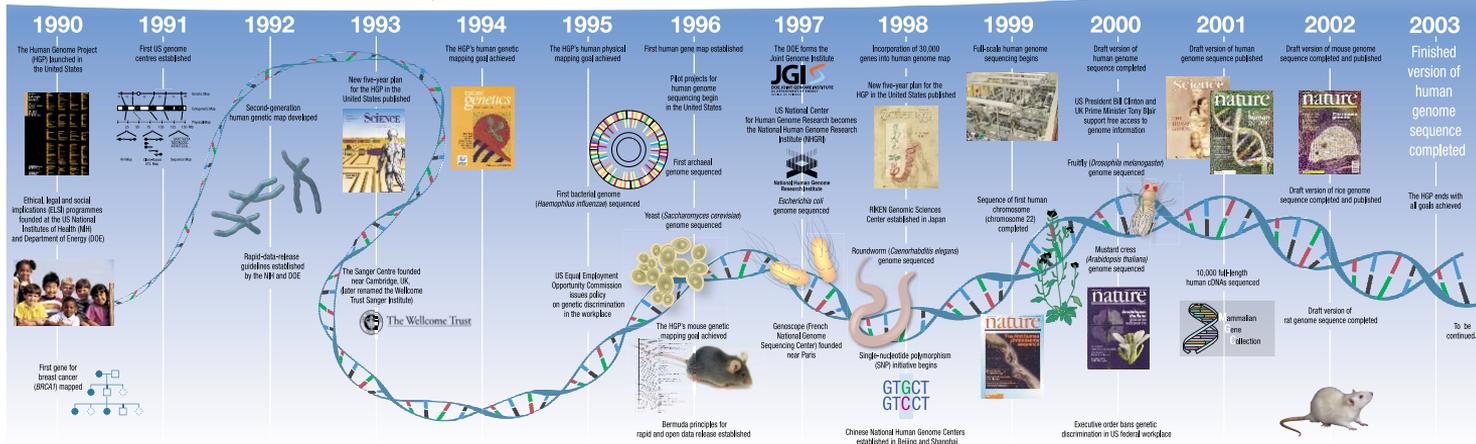
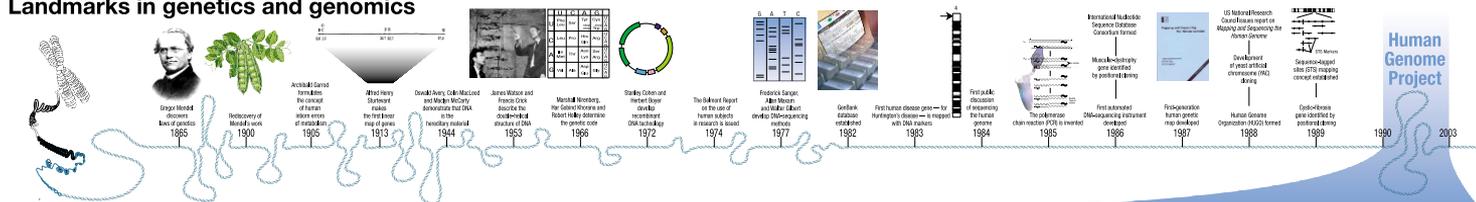
- Метод Сэнгера, dye-terminating sequencing, sequencing by synthesis, DNA sequencing by capillary electrophoresis
- Длина прочтений 700-1000 нк. Первые 15-40 нк – низкое качество.



Проект «Геном человека» Human Genome Project (HGP) 1990 - 2003

Рассчитан на 15 лет, с бюджетом \$3 млрд, завершён досрочно

Landmarks in genetics and genomics



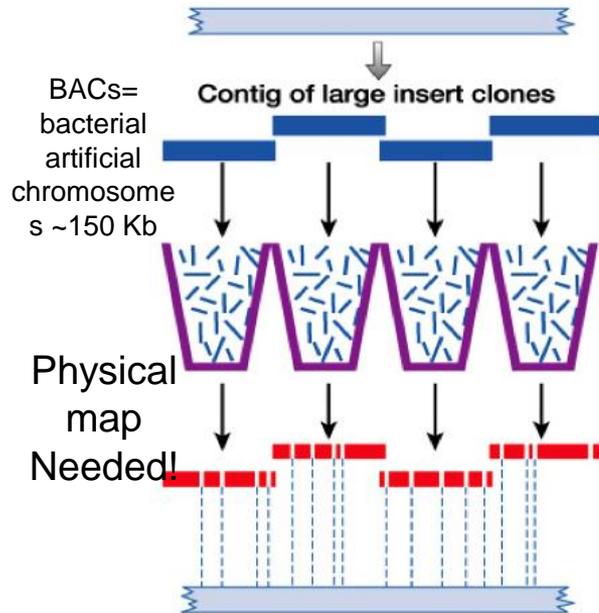
Проект геном человека: методы и подходы



F. Collins

1990

'BAC-by-BAC' approach
Hierarchical shotgun

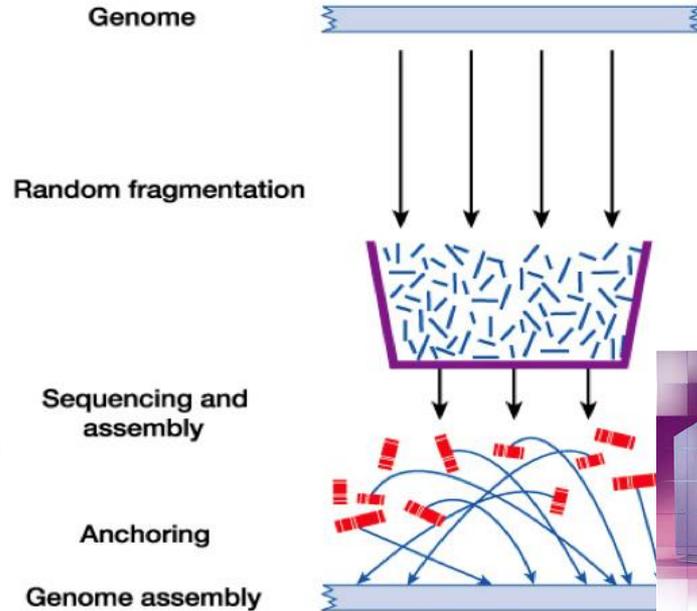


C. Venter

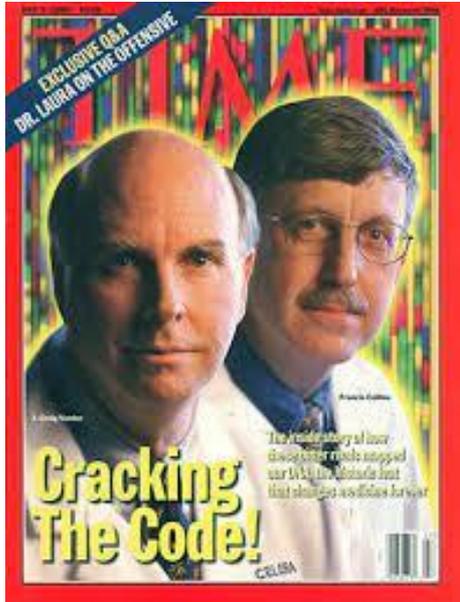
1998



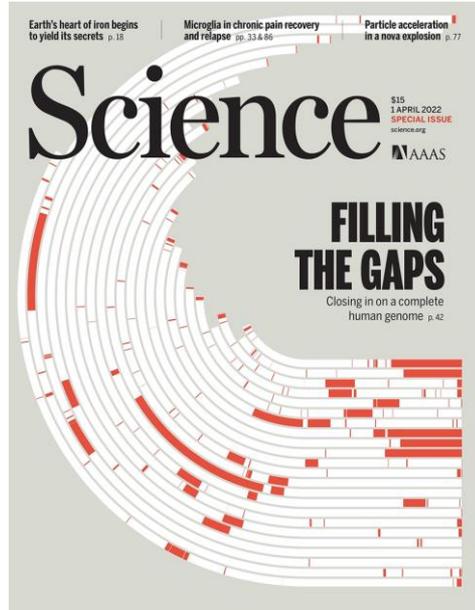
Whole-genome shotgun



2000



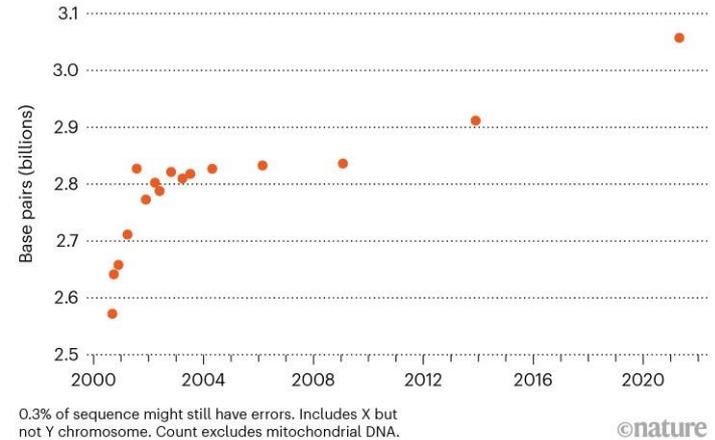
31 марта 2022



A Truly Complete Human Genome Sequence

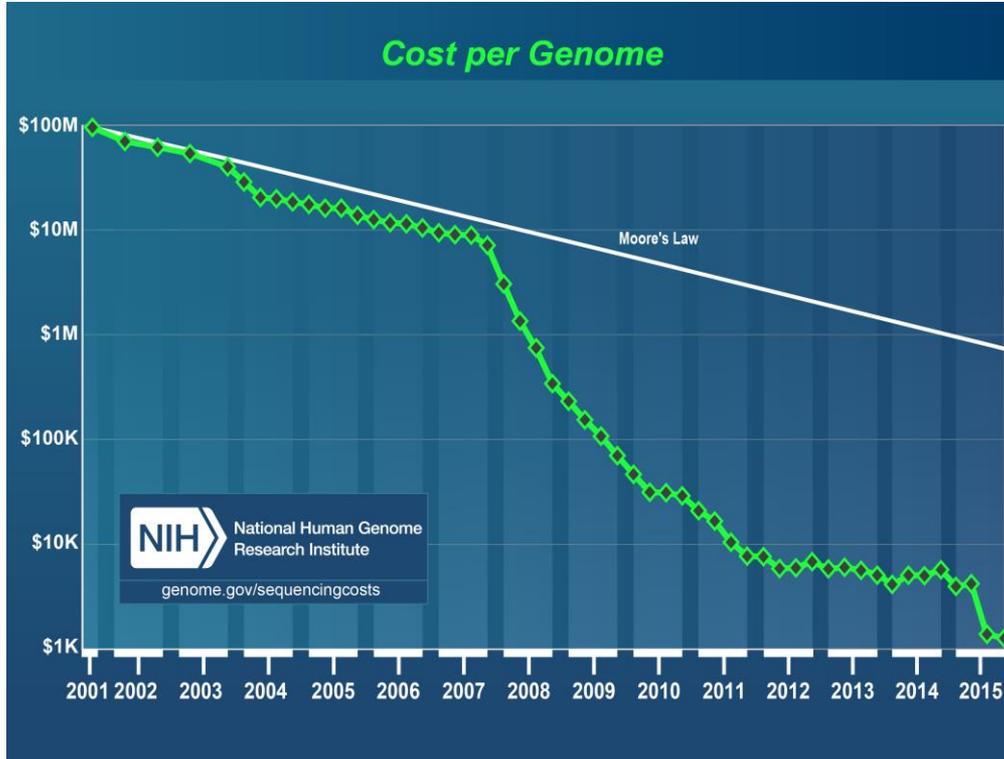
COMPLETING THE HUMAN GENOME

Researchers have been filling in incompletely sequenced parts of the human reference genome for 20 years, and have now almost finished it, with 3.05 billion DNA base pairs.



Human genome size ~ 3,117 billion base pairs (bp)

NGS sequencing



LETTERS

The complete genome of an individual by massively parallel DNA sequencing

David A. Wheeler^{1*}, Maithreyan Srinivasan^{2*}, Michael Egholm^{3*}, Yufeng Shen^{1*}, Lei Chen¹, Amy McGuire³, Wen He³, Yi-Ju Chen², Vinod Makhijani², G. Thomas Roth², Xavier Gomes², Karrie Tartaro^{2†}, Faheem Niazi², Cynthia L. Turcotte², Gerard P. Irzyk², James R. Lupski^{4,5,6}, Craig Chinault⁴, Xing-zhi Song¹, Yue Liu¹, Ye Yuan¹, Lynne Nazareth¹, Xiang Qin¹, Donna M. Muzny¹, Marcel Margulies², George M. Weinstock^{1,4}, Richard A. Gibbs^{1,4} & Jonathan M. Rothberg^{2†}

The association of genetic variation with disease and drug response, and improvements in nucleic acid technologies, have given great optimism for the impact of 'genomic medicine'. However,

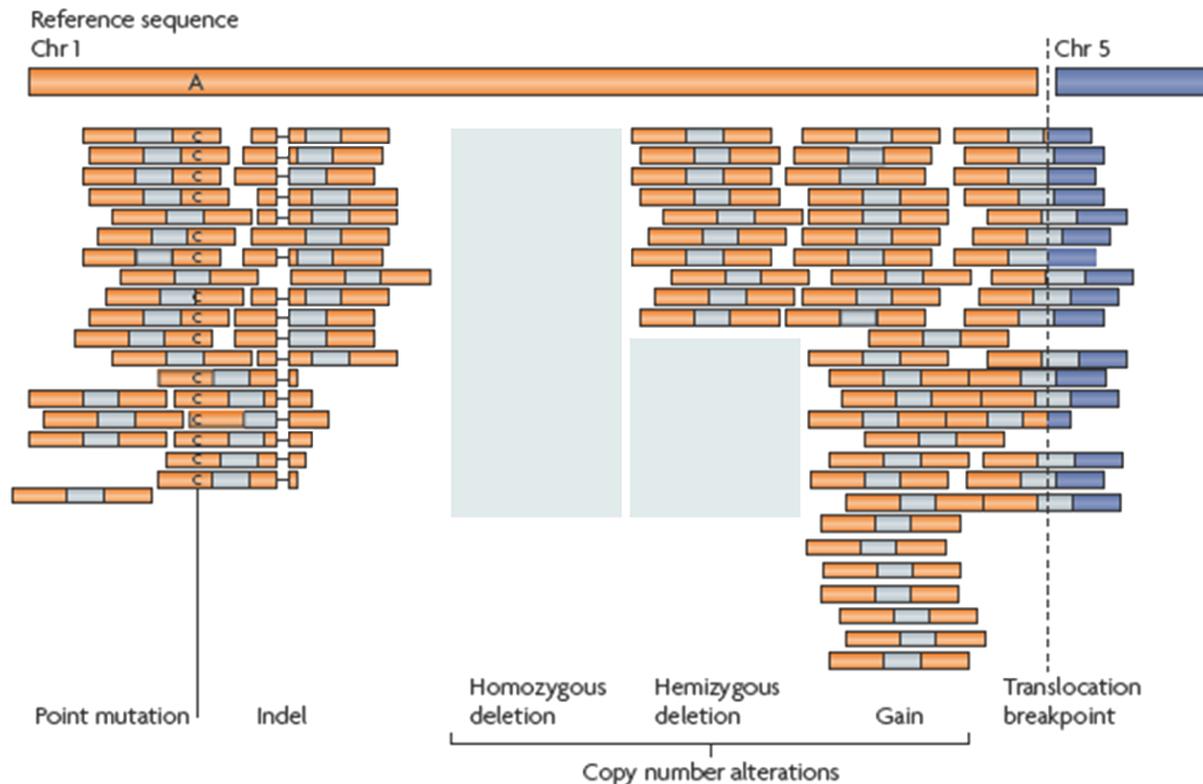
subject's DNA, including single nucleotide polymorphisms (SNPs), small insertions and deletions (indels), and copy number variation (CNV).

James Watson's genome sequenced at high speed



2008
454 Life Sciences
4 months
\$1.5 mln

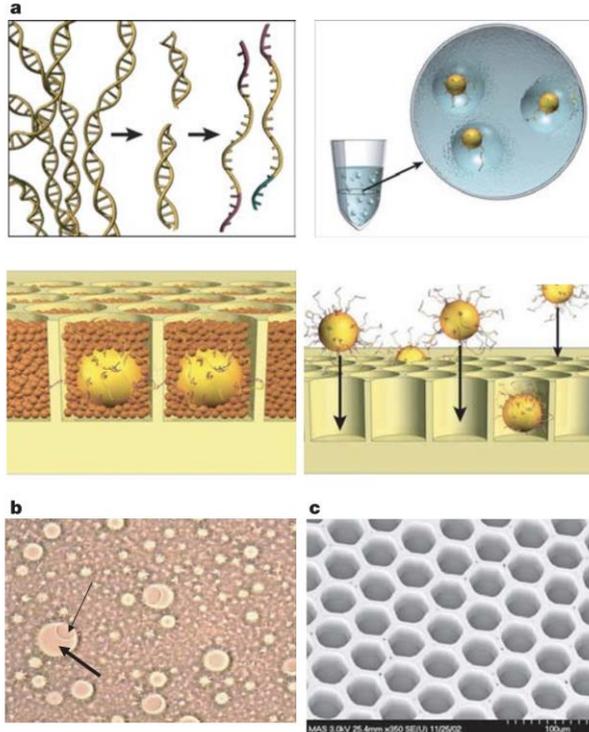
Ресеквенирование



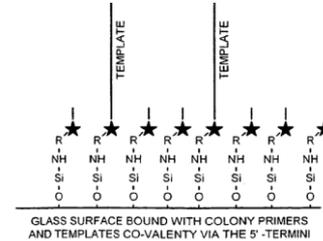
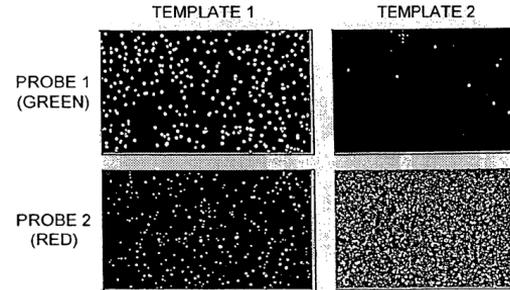
Nature Reviews Genet. 2010 Oct ;11(10):685-96.

Для достоверности в медицинских целях 30-100-кратное покрытие,
30-100-кратная глубина секвенирования

Технологии микрореакторов, массивно-параллельной работы с клональными библиотеками ДНК, «колониями» ДНК



SIMULTANEOUS DETECTION OF TWO INDEPENDENT DNA COLONIES



(12) **United States Patent**
Adessi et al.

(10) **Patent No.:** US 7,115,400 B1
(45) **Date of Patent:** *Oct. 3, 2006

(54) **METHODS OF NUCLEIC ACID AMPLIFICATION AND SEQUENCING**

(56) **References Cited**

(75) Inventors: **Celine Adessi**, Ambilly (FR); **Eric Kawashima**, Nyon (CH); **Pascal Mayer**, Eloise (FR); **Jean-Jacques Mermoud**, Geneva (CH); **Gerardo Turcatti**, Geneva (CH)

U.S. PATENT DOCUMENTS
5,514,539 A * 5/1996 Bukh et al. 435/5

FOREIGN PATENT DOCUMENTS
EP 0543484 A2 5/1993

(73) Assignee: **Solexa Ltd.**, Essex (GB)

(Continued)

Технологии секвенирования нового (второго) поколения
NGS – next generation sequencing
Массивно-параллельное секвенирование
Massively parallel sequencing

Sequencing by synthesis

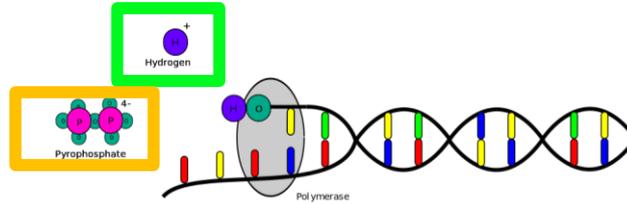
- Пиросеквенирование – по технологии компании 454 Life Sciences/Roche
– Первая технология на рынке. С 2016 года не поддерживается.
- Секвенирование на основе обратимой терминации синтеза ДНК– по технологии компаний Solexa/Illumina.
– 90% всего секвенирования выполняется сейчас на этой технологии.
- Полупроводниковое секвенирование - Ion Torrent
- DNBSEQ метод от компании MGI на основе ДНК-наношаров (CoolMPS версия)

Sequencing by ligation

- SOLiD от Applied Biosystems. Не поддерживается
- DNBSEQ метод от компании MGI на основе ДНК-наношаров (cPAS версия)

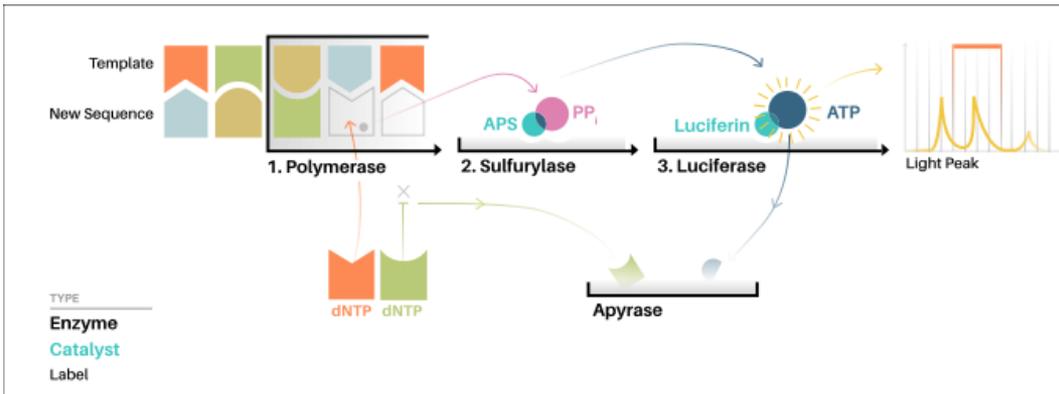
Sequencing by binding (PacBio announced)

Пиросеквенирование/ Ионное полупроводниковое секвенирование

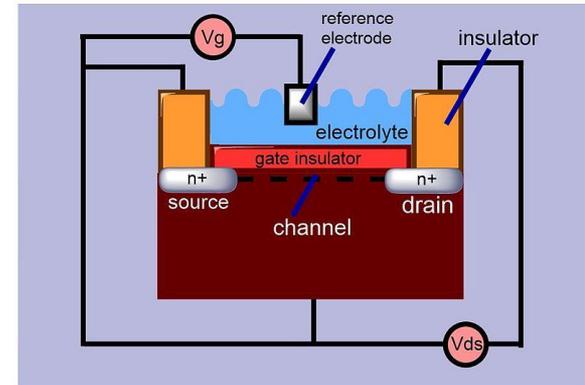


Hydrogen and pyrophosphate are released.

Пирозфосфат ддетектируется через активность
люциферазы



ion-sensitive field-effect transistor (ISFET)
Ион чувствительный полевой транзистор
рН-чувствительный транзистор



Пиросеквенирование от 454 Life Sciences

- 2005 год – первый секвенатор (GS20 – 25 млн. нт за прогон)
 - 2005 секвенированы геномы *Mycoplasma* и *Streptococcus*
 - 2007 год – улучшенный секвенатор 454 GS-FLX
 - 2008 – секвенирован геном James Watson
 - Компания 454 LS куплена Roche в 2007 году.
 - 2012 – FLX+
 - 2006 – 2013 - ... – секвенирование генома неандертальца
 - С 2016 года технология закрыта, неконкурентноспособна (по сравнению с Illumina, IonTorrent)
-
- FLX+: 1 млн прочтений, до 700 нт, 0.7 Гб, \$8К, 10 часов за прогон
 - Хорошая точность
 - Ошибки в гомополимерных повторах





Применение:

- секвенирование de novo коротких фрагментов ДНК (180 п.о.);
- ресеквенирование любого участка генома человека, модельных объектов, микроорганизмов или вирусов;
- количественный анализ частот аллельной встречаемости;
- мутационный анализ: идентификация точечных однонуклеотидных замен (SNP), вставок и делеций;
- количественный анализ метилирования ДНК (может быть совмещен с SNP типированием);
- верификация и валидация результатов полногеномного анализа.

Технические характеристики:

- производительность — от 1 до 48 образцов;
- пробоподготовка — автоматизированная на самом приборе;
- сенсорный дисплей управления;
- размеры, Ш × Г × В, см — 25 × 30 × 30;
- вес, кг — 8,5.

Полупроводниковое секвенирование от Ion Torrent

- 2007 год – Ion Torrent Systems – основана компания
- 2010/11 - первый секвенатор Personal Genome Machine (PGM)
- 2010 - Компания приобретена Life Technologies, позднее ThermoFisher
- 2012 - секвенатор Ion Proton (в 1000 раз мощнее PGM), геном человека за \$1000 (не за один прогон)



- Длина ридов до 400 нт, высокая скорость, нет сложной химии, дешевизна прибора,
- Позиционируется для задач диагностики – каждому доктору в кабинет
- Ошибки в гомополимерных повторах, нет возможности парноконцевого секвенирования





Ion GeneStudio S5 Systems



Ion Torrent Genexus System

- Быстрое время от эксперимента до результата
- Высокий уровень автоматизации
- Хорош для медицинских исследований
- Чипы дают ограниченное число прочтений

SOLiD (Sequencing by Oligonucleotide Ligation and Detection) от ABI

- 2003 – polony sequencing technology (G. Church)
- 2006/08 год – первый секвенатор по технологии SOLiD
- 2015 ... - сняты с производства
- 2017 - закончена техподдержка
- Короткие риды (50-75 нт), очень дешево (\$0.13/млн нт)
- Высокая точность 99.94%. Использовался для ресеквенирования, анализа вариаций
- Медленно – 1 запуск – до 14 дней
- Нет проблем с гомополимерными повторами
- Проблемы с палиндромными сиквенсами

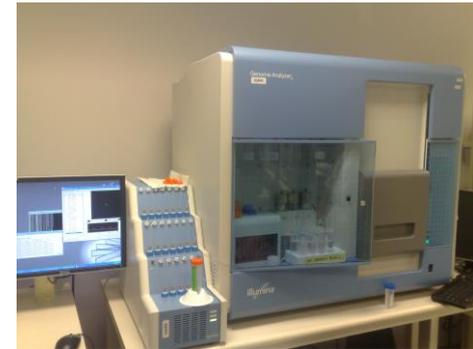


Applied Biosystems™

5500xl Genetic Analyzer

NGS платформа Illumina/Solexa

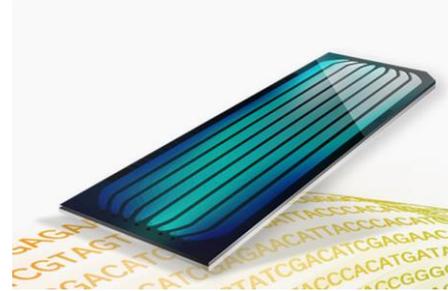
- 1998: Shankar Balasubramanian, David Klenerman (Cambridge) founded Solexa company
- 2005 секвенирован геном phiX-174
- 2006 – первый секвенатор (Genome Analyzer – 1 млрд. нт. за прогон)
- 2007 год – acquired by Illumina
- 2007 - ... - эволюция приборов и технологии. Технология на четырех флюоресцентных метках, на двух, на одной + полупроводниковая детекция
- Линейка секвенаторов
- Длина прочтений: до 150-300 нт.
- Возможность прочтения прямой и обратной цепей (**парноконцевое секвенирование, paired-end sequencing**)



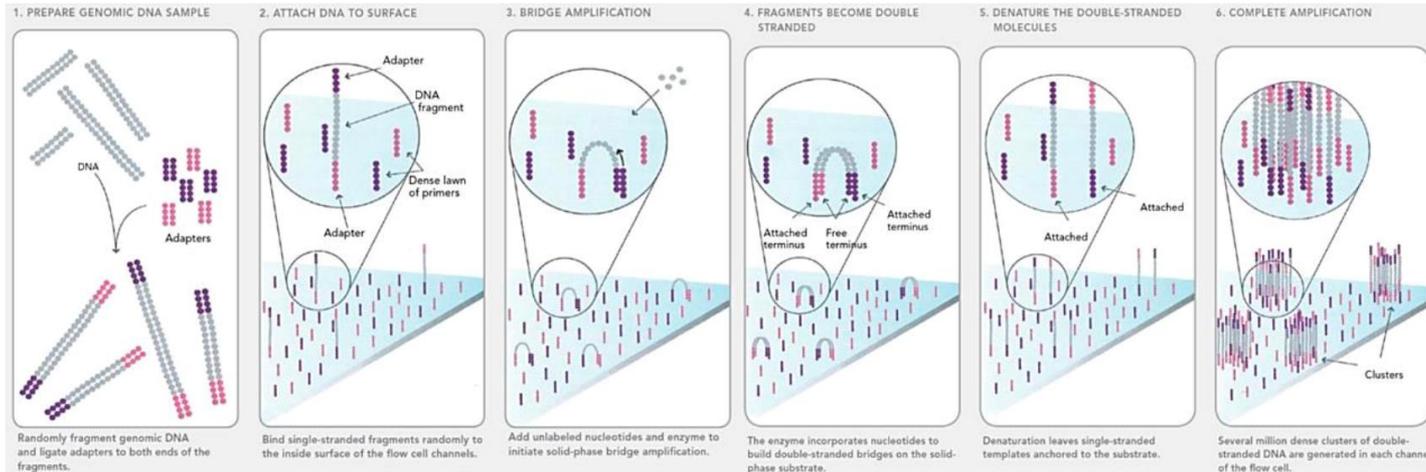
Genome Analyzer

NGS платформа Illumina/Solexa

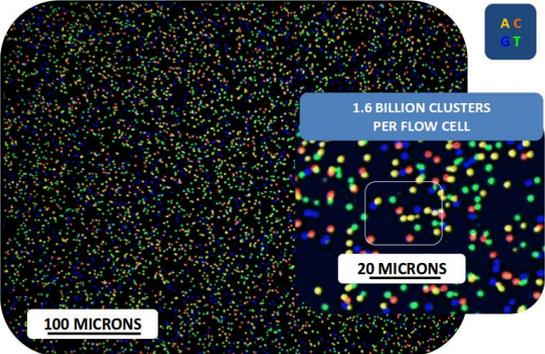
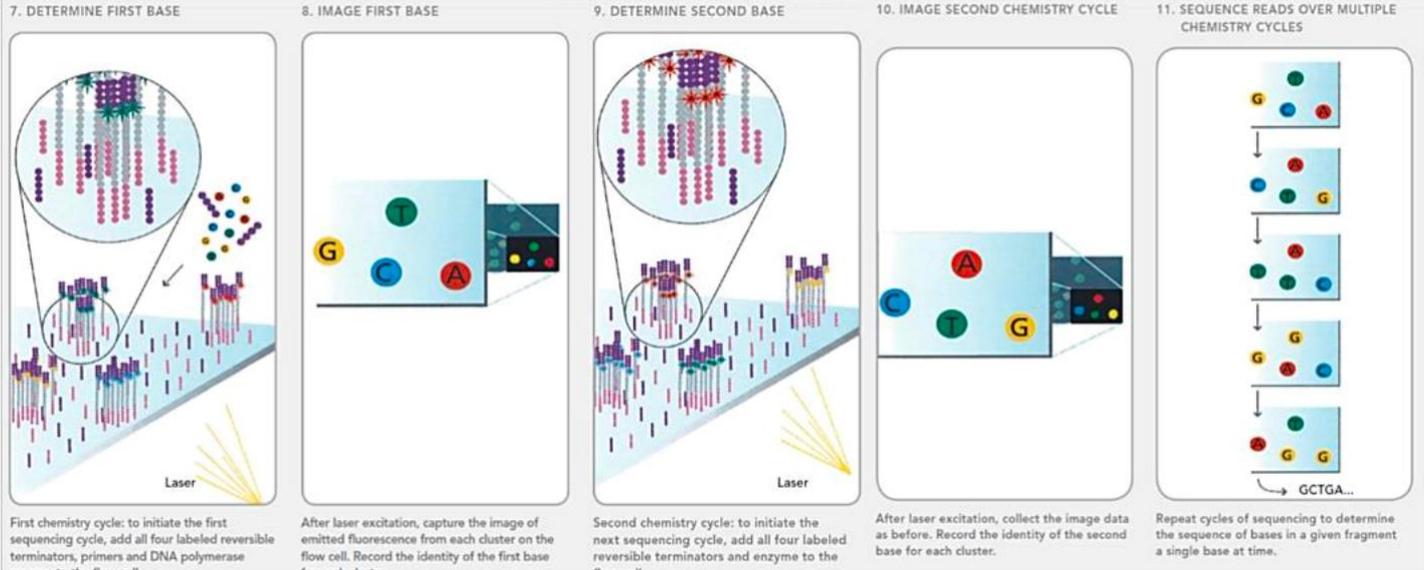
Sequencing by synthesis
Bridge amplification
Dye terminator sequencing



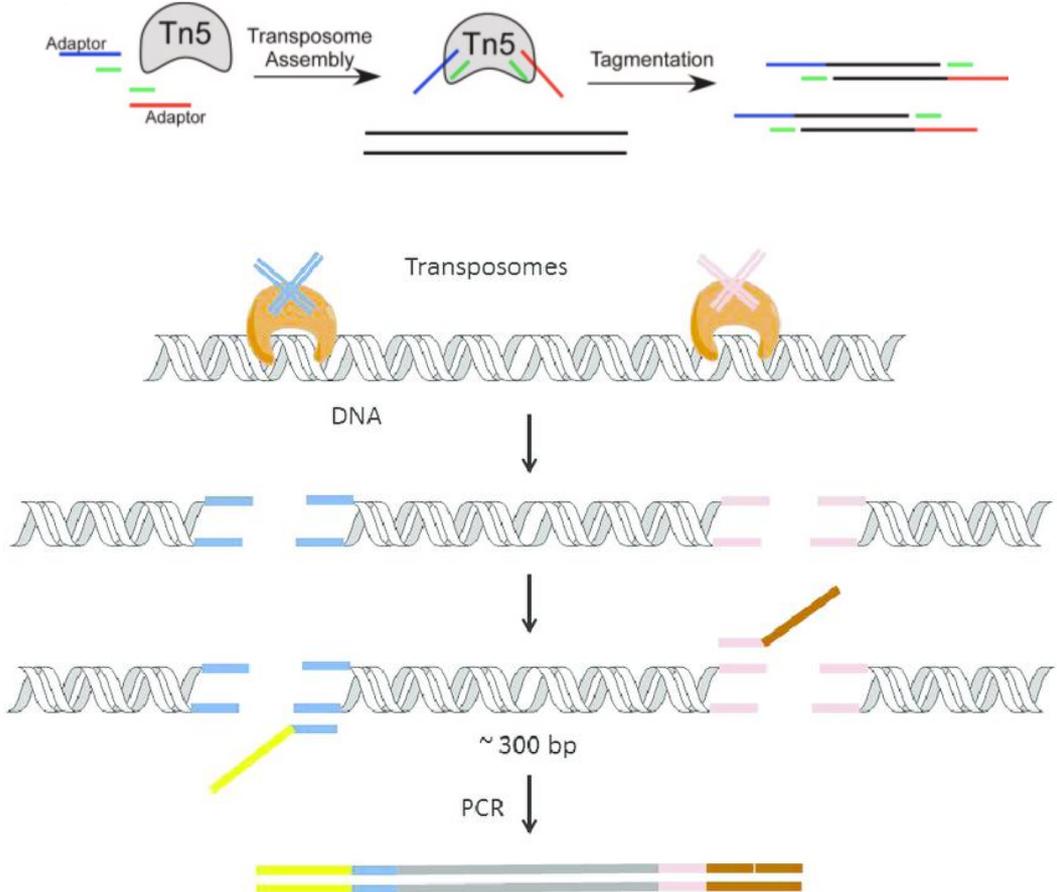
Flow cell
Проточная ячейка



NGS платформа Illumina/Solexa



Подготовка библиотеки (Illumina library prep)



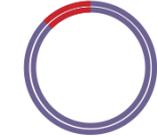
Технология DNBSEQ от MGI

- 1999 – основан Beijing Genomic Institute => BGI Group
 - 2009 – Complete Genomics отсеквенировала геном человека с помощью своей проприетарной технологии (статья в Science)
 - 2013 – BGI приобретена компания Complete Genomics
 - 2014 – BGISEQ-1000 первый секвенатор
 - 2014 - ... дальнейшее развитие технологии
-
- Позиционируется как альтернатива Illumina, которая дешевле и быстрее
 - Длина ридов до 150 нт
 - Парноконцевое секвенирование
 - performances of the MGISEQ-2000 and DNBSEQ-T7 platforms are comparable to that of the Illumina NovaSeq 6000



Library generation through RCA

d In-solution DNA nanoball generation (Complete Genomics (BCI))

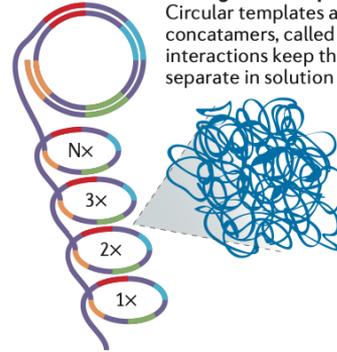


Adapter ligation
One set of adapters is ligated to either end of a DNA template, followed by template circularization

Cleavage
Circular DNA templates are cleaved downstream of the adapter sequence

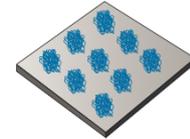


Iterative ligation
Three additional rounds of ligation, circularization and cleavage generate a circular template with four different adapters

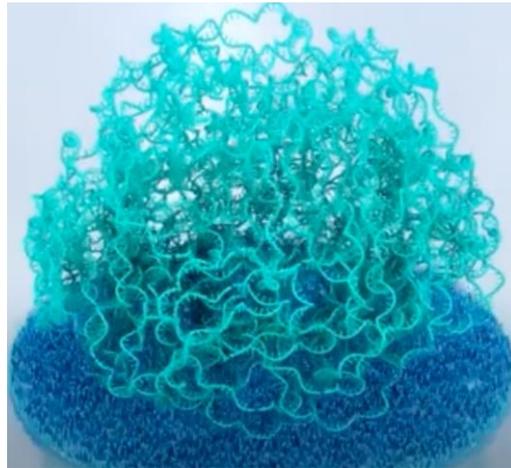


Rolling circle amplification

Circular templates are amplified to generate long concatamers, called DNA nanoballs; intermolecular interactions keep the nanoballs cohesive and separate in solution



Hybridization
DNA nanoballs are immobilized on a patterned flow cell



DNA nanoball

CoolIMPS

sequencing by synthesis

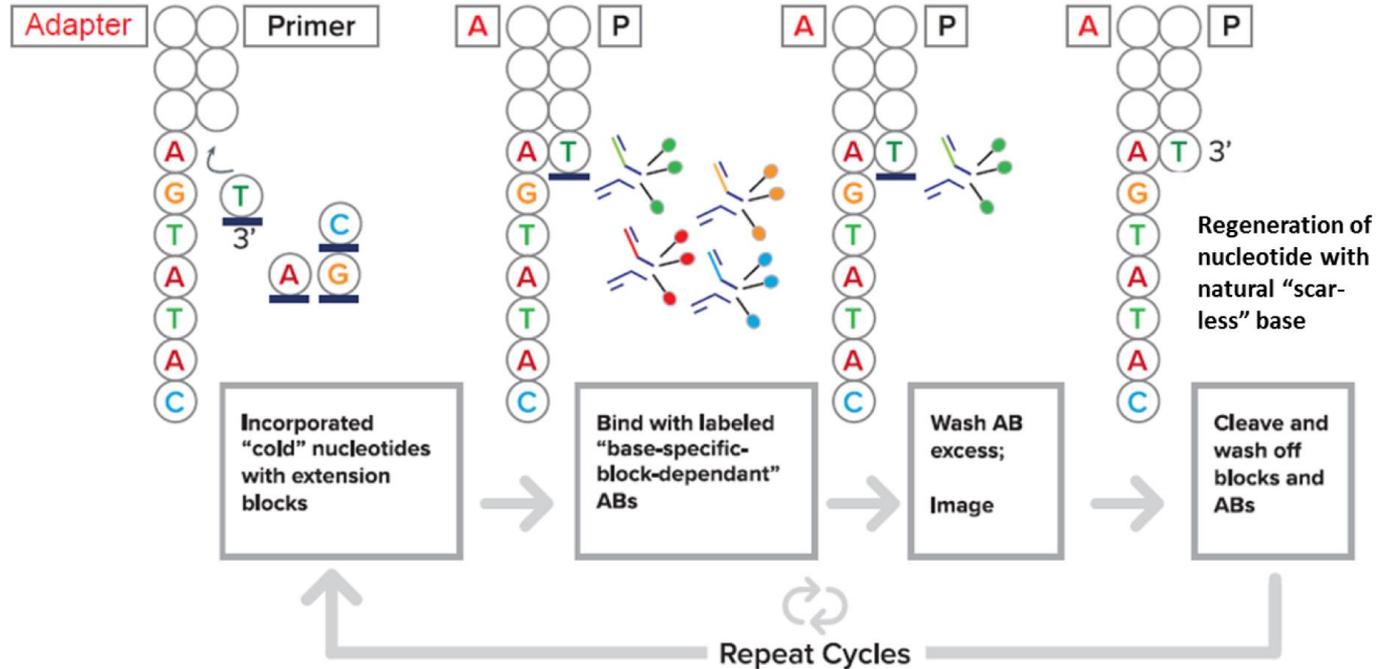


Figure 1: CoolIMPS™ process overview. Bars (—) on the unlabeled ("cold") nucleotides depict removable 3' chemical blocks. Antibodies specific for RTs with natural nucleobase are depicted with three dye molecules to increase fluorescent signal.



Sequencing by binding

2021: Компания PacBio приобрела Omniome
Сверхвысокая точность (Q40?)
2022 октябрь – коммерческий продукт



- https://youtu.be/i_mSaNBOVmQ

Методы секвенирования третьего поколения

- Плюсы:

- 1) Можно секвенировать длинные фрагменты ДНК
- 2) Можно получать длинные прочтения (до 20 000 нт)
- 3) Не обязательно амплифицировать для считывания сигнала => более простая пробоподготовка
=> портативные секвенаторы
- 4) Эпигенетическая разметка ДНК сохраняется (NanoPore)

- Минусы:

- 1) Слабый сигнал, высокий процент ошибок

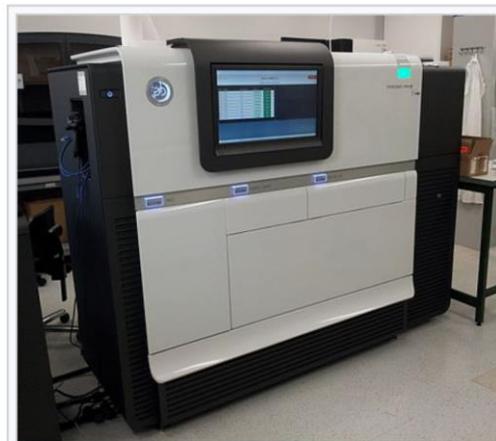
- Технологии:

- 1) SMRT by Pacific Biosciences
- 2) Oxford Nanopore
- 3) Illumina Infinity 2022?

Технология PacBio



- 2004 – основана Pacific Biosciences
 - 2010 – первый прибор PacBio RS
 - 2013 - PacBio RS II
 - 2015 - Sequel System
 - 2019 - Sequel II System
 - 2020 - Sequel IIe System
 - 2022 – Revio System
-
- Определение структурных вариаций генома
 - Может секвенировать эпигенетические модификации
 - Может секвенировать РНК напрямую.

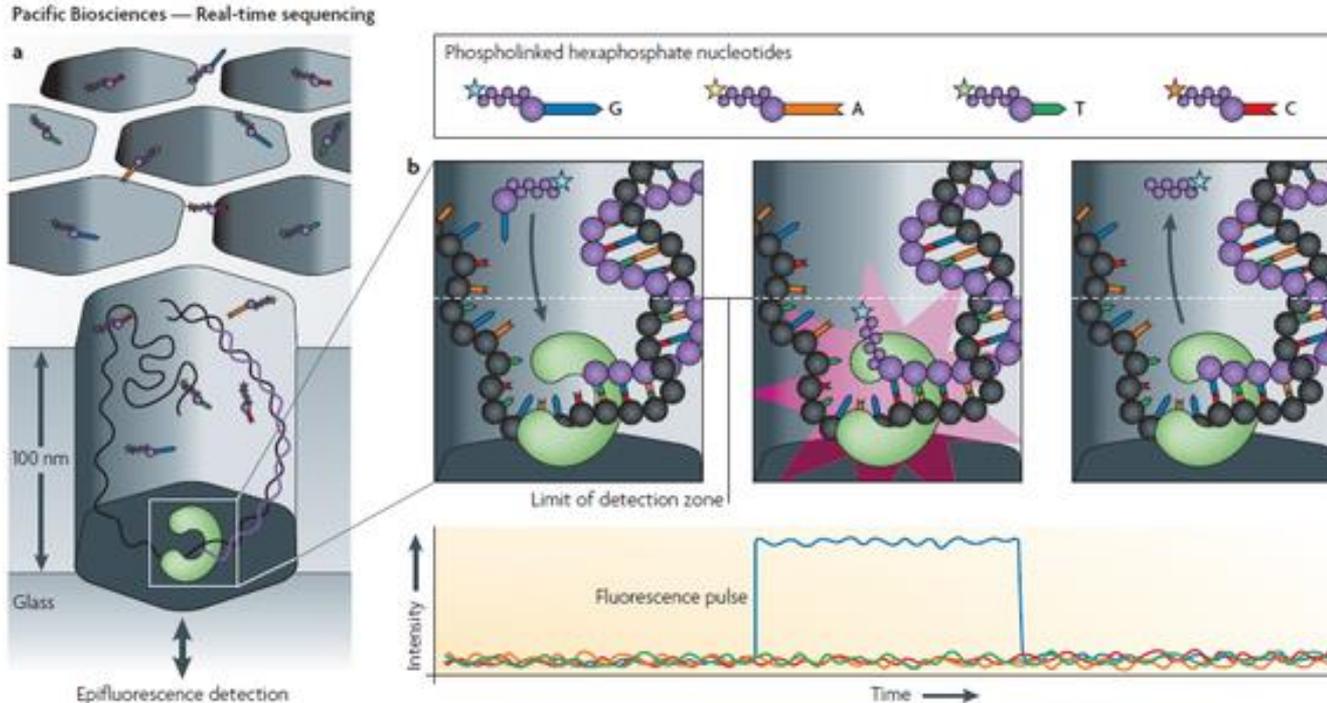


A PacBio RSII sequencer



Технология PacBio

Single-molecule real-time (SMRT) sequencing



https://youtu.be/_ID8JyAbwEo

<https://youtu.be/NHCJ8PtYCFc>

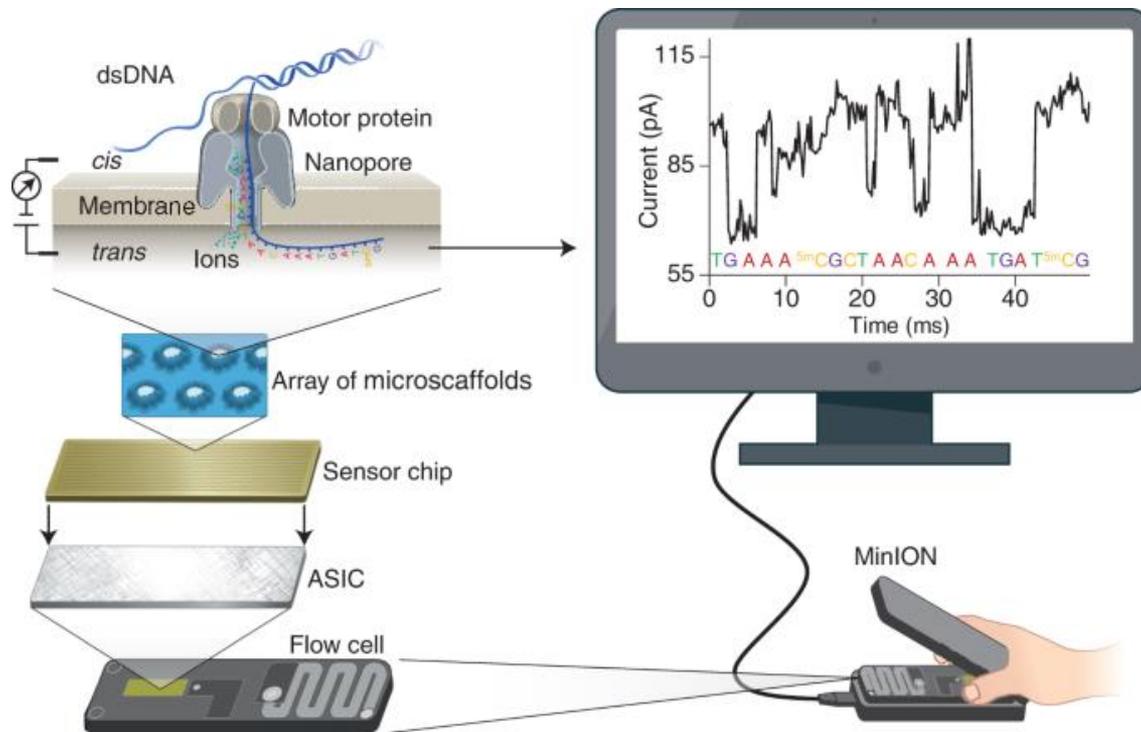
Технология Oxford Nanopore

- 2005 – основана Oxford Nanopore Technologies
- 2014 – первый прибор MinION

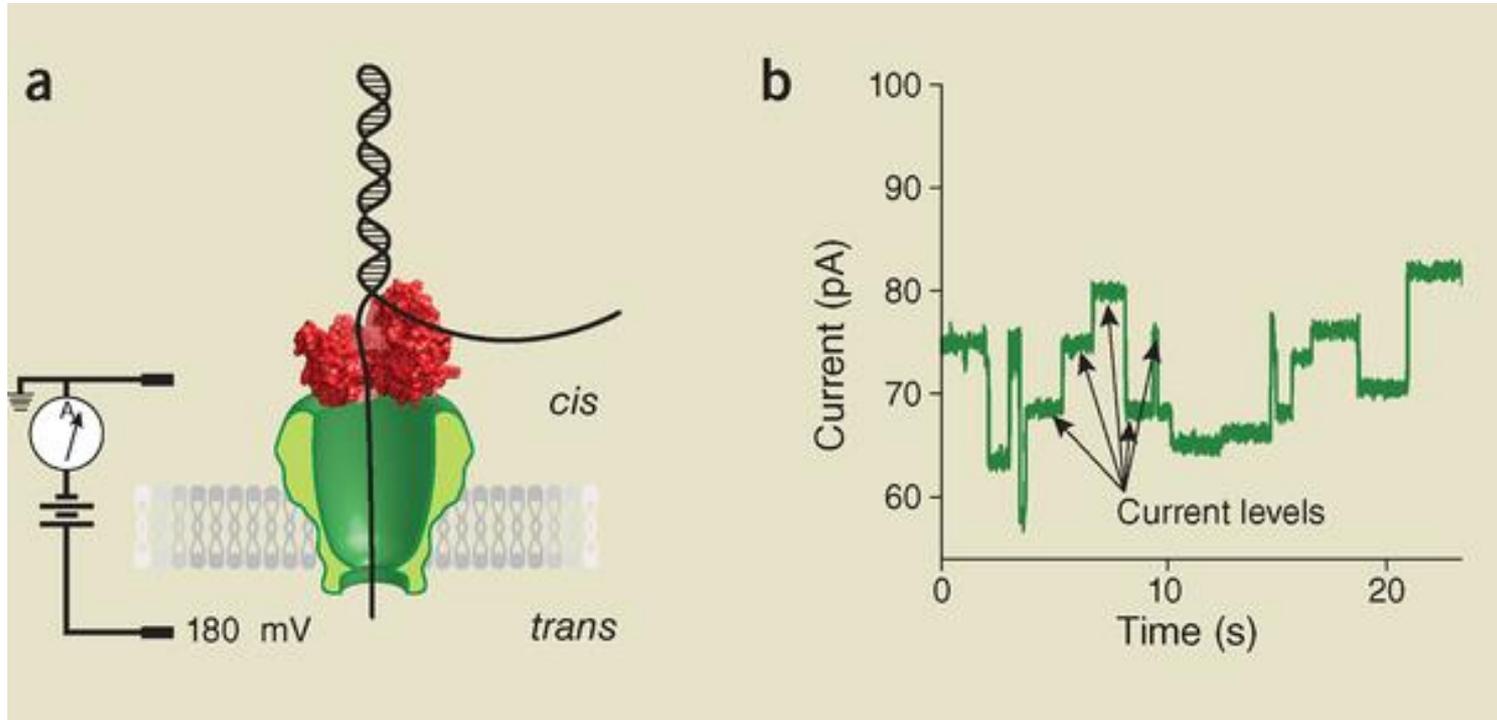


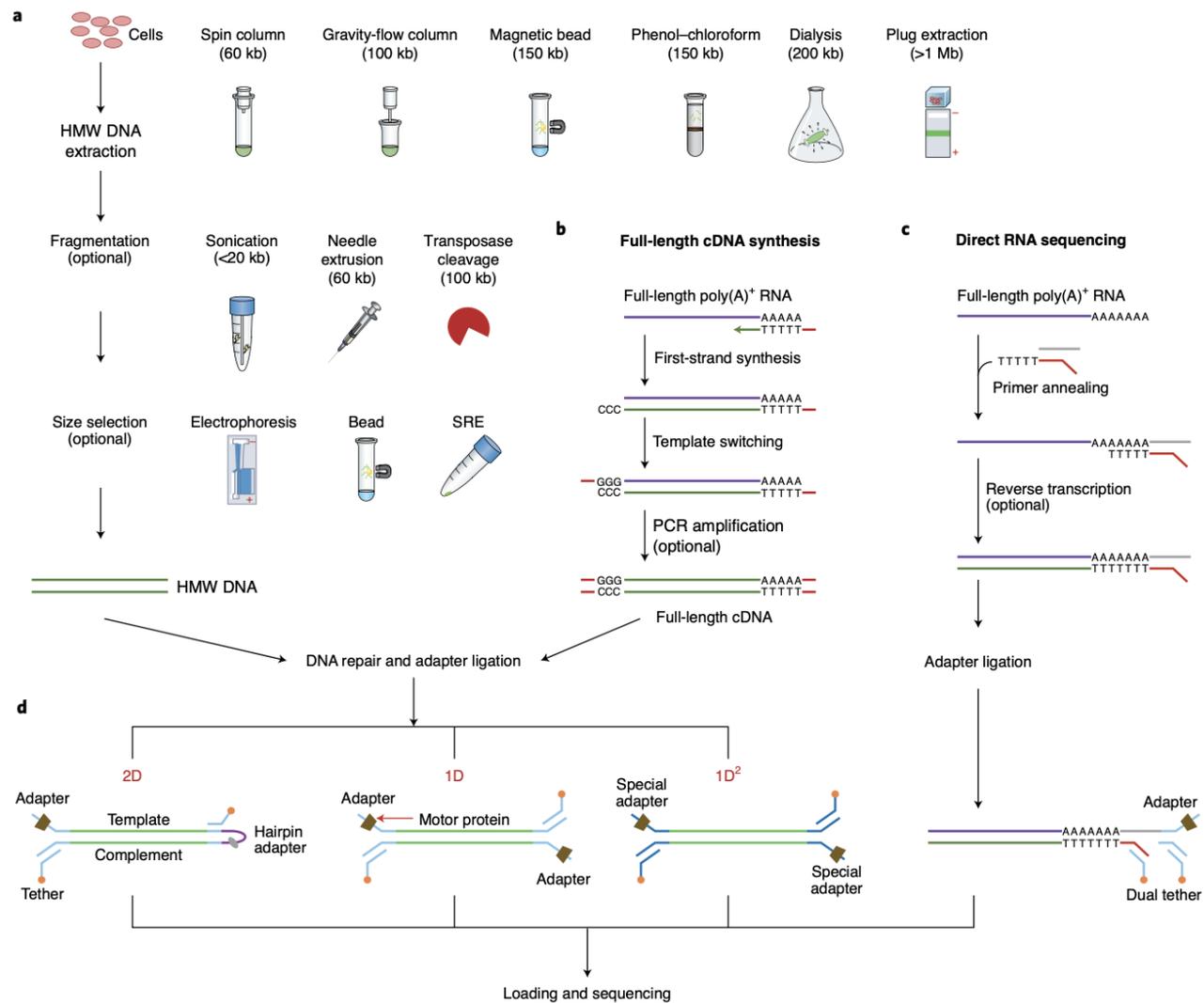
- Портативный прибор
- Точность 99,9%
- Определение структурных вариаций генома
- Может секвенировать эпигенетические модификации
- Может секвенировать РНК напрямую.

Технология Oxford Nanopore



<https://youtu.be/RcP85JHLmnl>





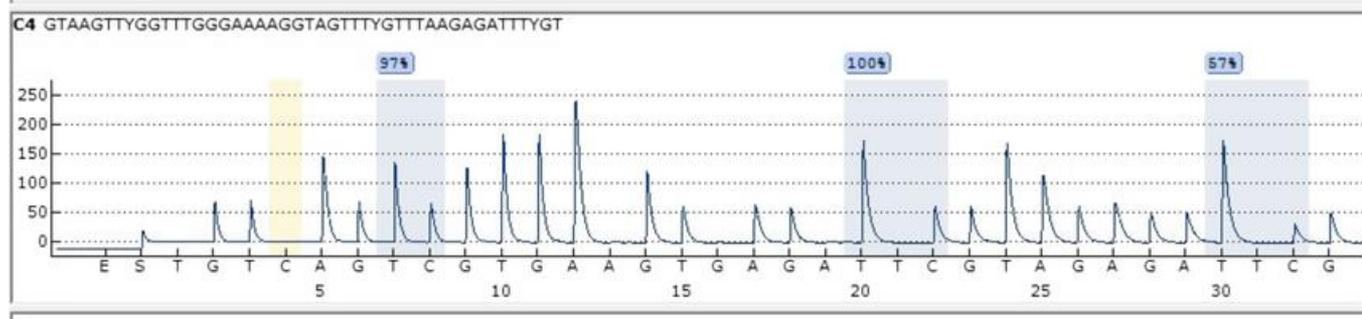
	Flongle	MinION	GridION (5 flow cells)	PromethION (48 flow cells)
				
Maximum run time	16 hours	72 hours	72 hours	64 hours
Theoretical 1D maximum yield	Up to 3.3 Gb	Up to 40 Gb	Up to 200 Gb	Up to 15 Tb
Current 1D maximum yield	Up to 2 Gb	Up to 30 Gb	Up to 150 Gb	Up to 8.6 Tb
Available channels	Up to 126	Up to 512	Up to 2,560	Up to 144,000

The sequencing players at a glance

Company	Type of tech	Length of read	Technology details
Illumina	Short read	~500 bases	SBS-based
Illumina	Long read	~10,000 bases	Infinity: in early research
Oxford Nanopore	Long read	Up to 1 million bases, usually 5,000-20,000	Nanopore-based
Pacbio	Long read	Up to 50,000 bases, usually ~25,000	SBS-based
Pacbio	Short read	~500 bases	SBB-based, acquired via Omniome

SBS=sequencing by synthesis. SBB=sequencing by binding. Source: interviews.

PyroMark



Кратко обсудим плюсы и минусы разных методов

Sequencing by synthesis

- Пиросеквенирование – по технологии компании 454 Life Sciences/Roche
– Первая технология на рынке. С 2016 года не поддерживается.
- Секвенирование на основе обратимой терминации синтеза ДНК– по технологии компаний Solexa/Illumina.
– 90% всего секвенирования выполняется сейчас на этой технологии.
- Полупроводниковое секвенирование - Ion Torrent
- DNBSEQ метод от компании MGI на основе ДНК-наношаров (CoolMPS версия)

Sequencing by ligation

- SOLiD от Applied Biosystems. Не поддерживается
- DNBSEQ метод от компании MGI на основе ДНК-наношаров (cPAS версия)

Sequencing by binding (PacBio)

Третье поколение

SMRT от PacBio

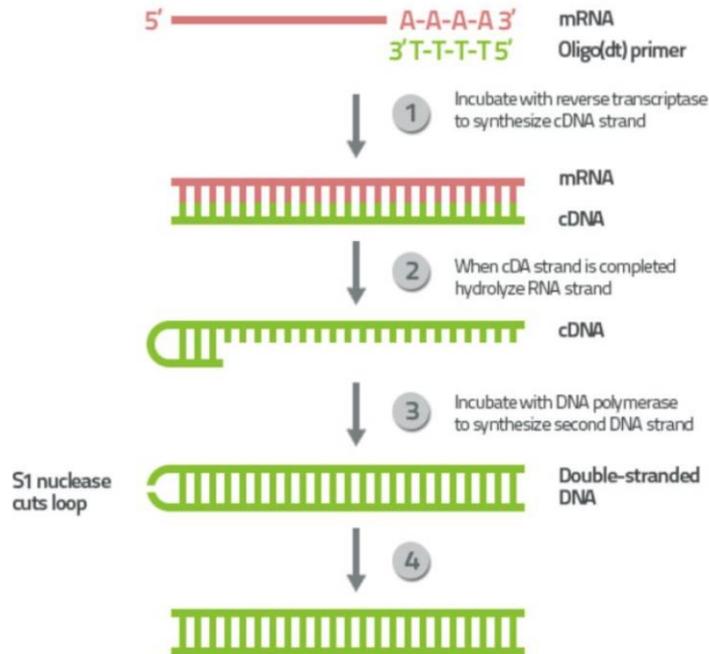
Nanopore Sequencing

Производные методы секвенирования

- RNA-seq
- Модификации ДНК (бисульфитная конверсия, NanoPore)
- ChIP-seq, ChIP-exo
- Доступность хроматина (DNase-seq, FAIR-seq, ATAC-seq)
- Positionирование нуклеосом (MNase-seq, Mnase-Exo III)
- Ribo-seq
- DAM-ID
- DAM-метилирование
- 3C-методы (HiC, Micro-C)
- ДНК-РНК
- Single cell

Секвенирование РНК

Комплементарная ДНК (кДНК, cDNA), кДНК библиотеки

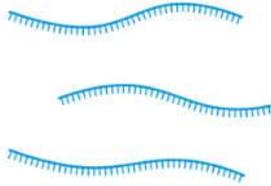


кДНК может быть секвенирована по Сэнгеру или NGS

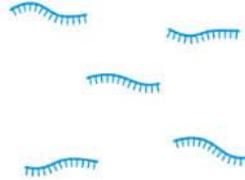
RNA-seq

RNA Sequencing

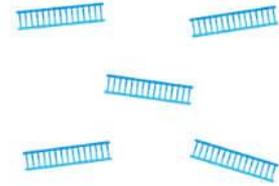
① Isolate RNA from samples



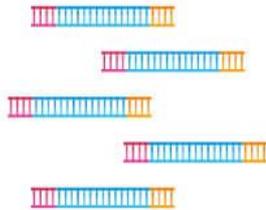
② Fragment RNA into short segments



③ Convert RNA fragments into cDNA



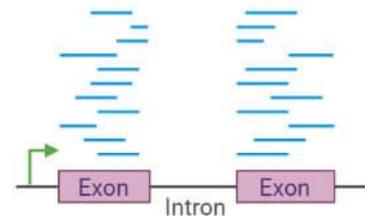
④ Ligate sequencing adapters and amplify



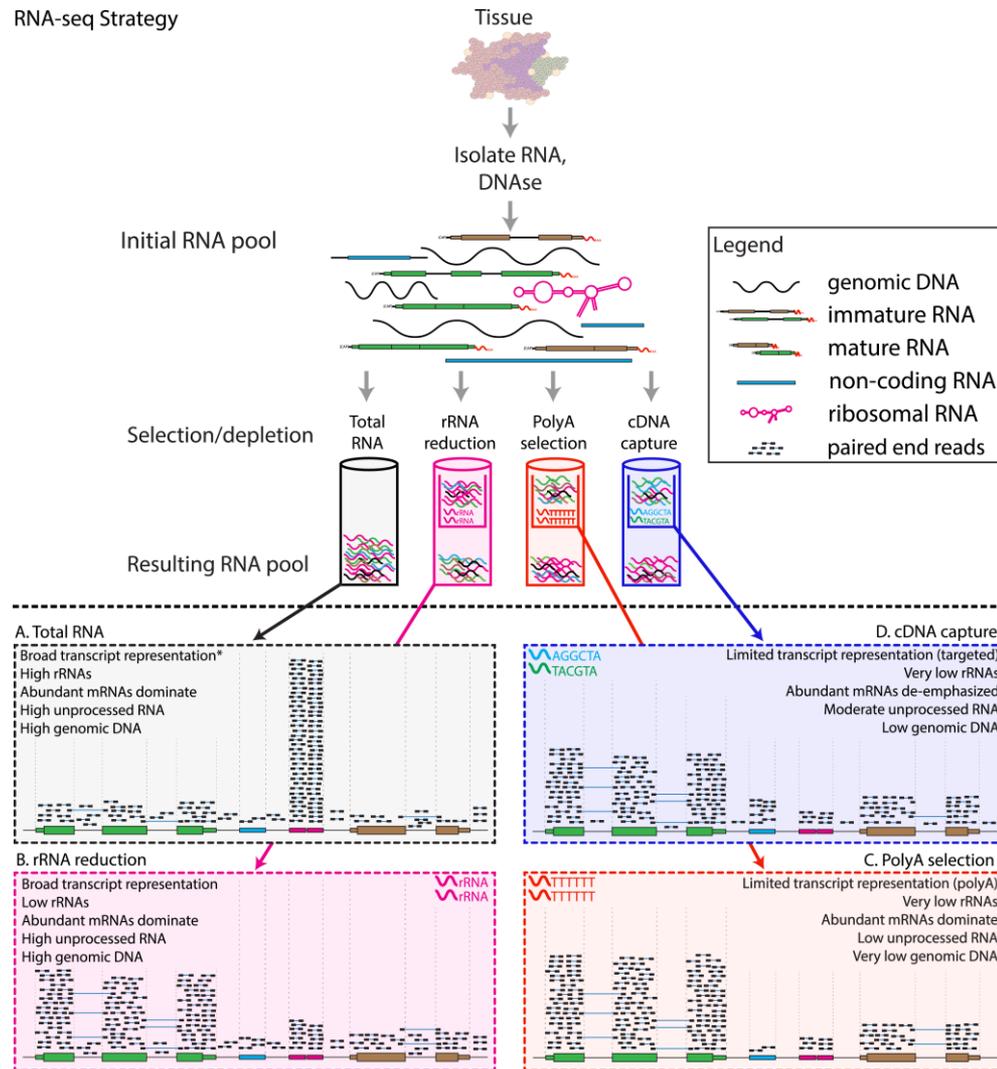
⑤ Perform NGS sequencing



⑥ Map sequencing reads to the transcriptome/genome

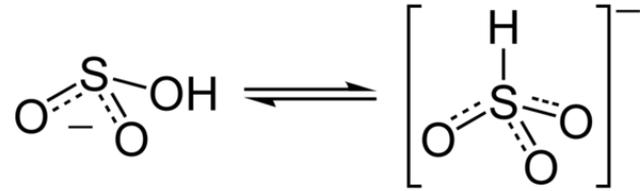
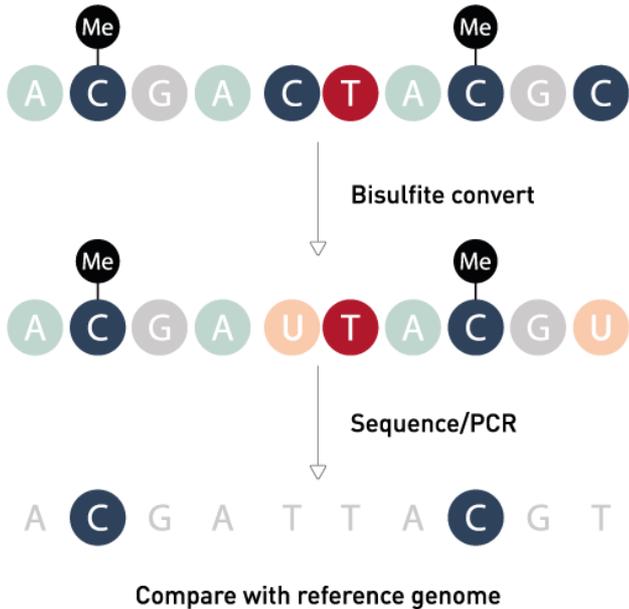


RNA-seq Strategy



Анализ модификаций ДНК

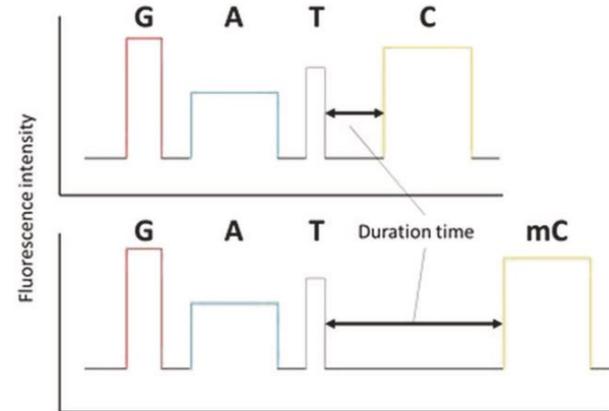
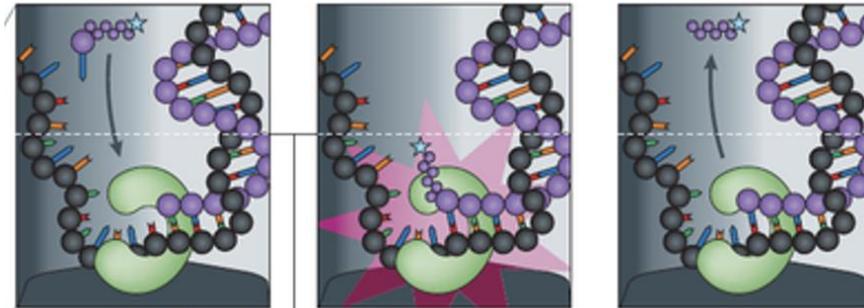
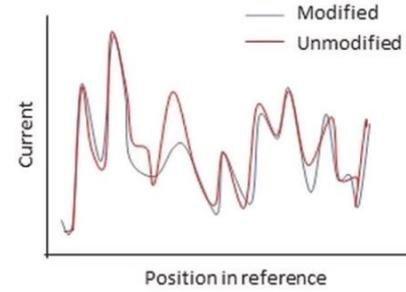
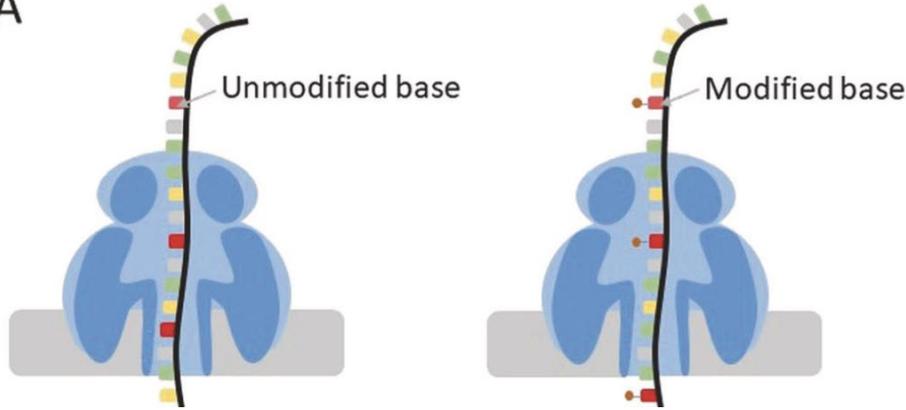
- Пример определение 5m-цитозинов в ДНК, бисульфитное секвенирование



Бисульфит (гидросульфит)

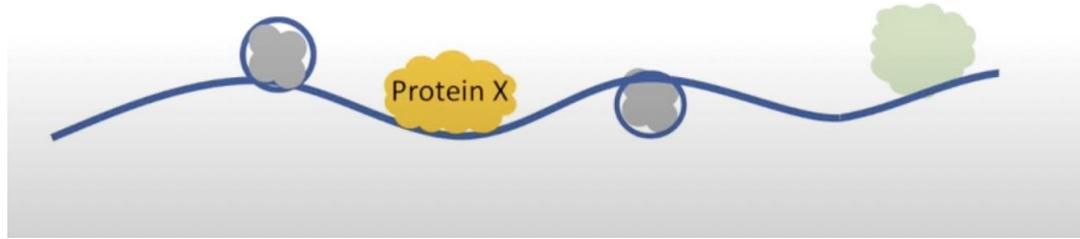
Oxford Nanopore и PacBio

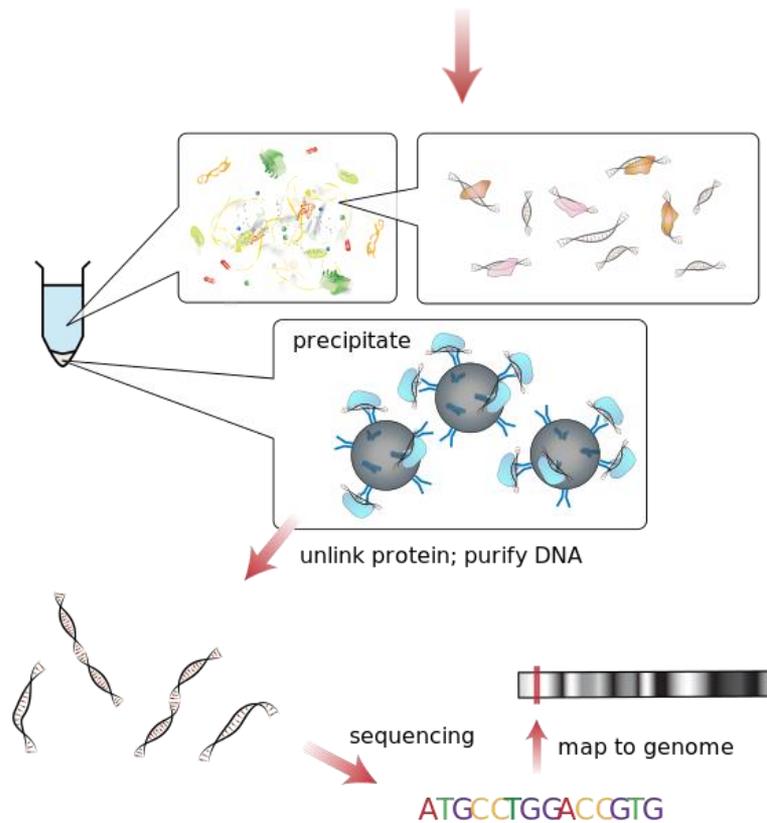
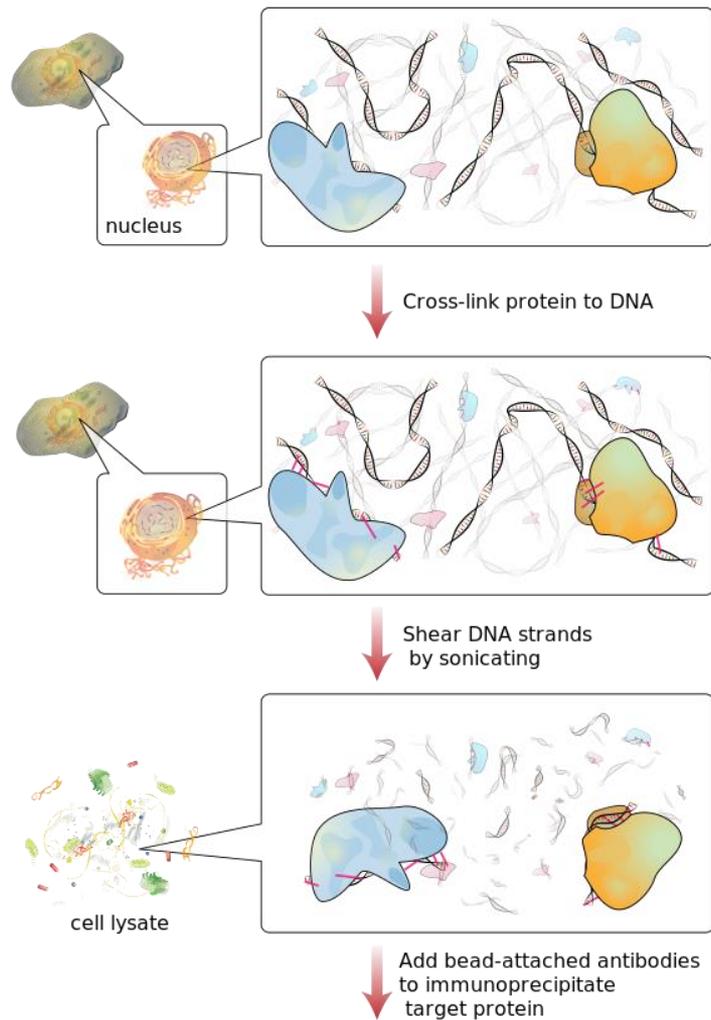
A



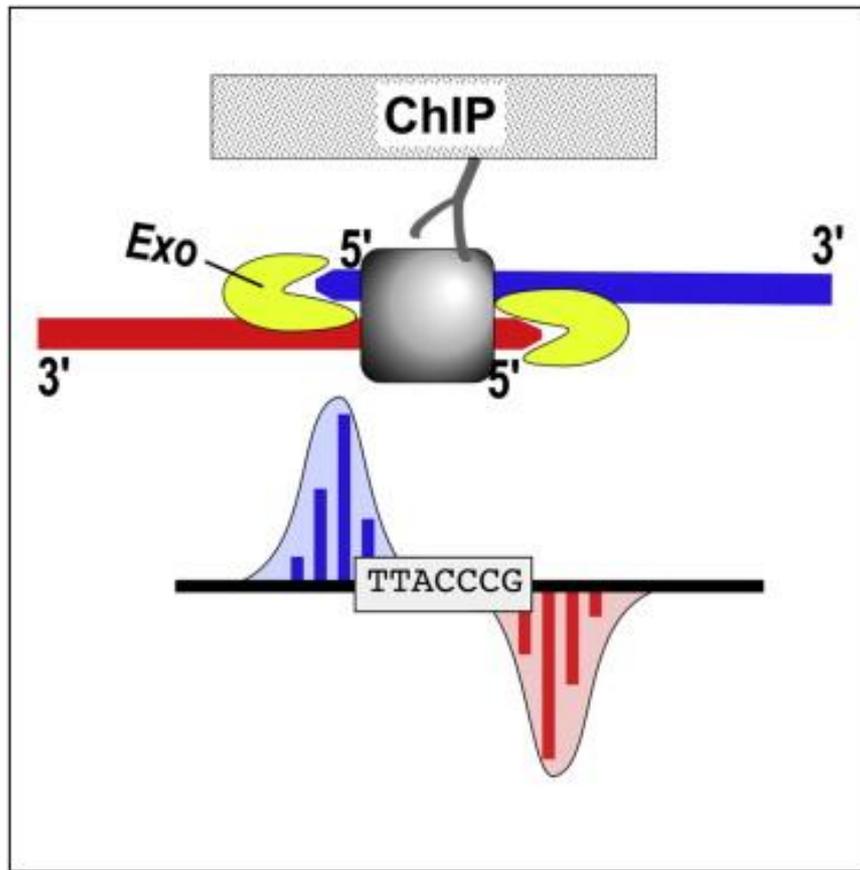
ChIP-seq

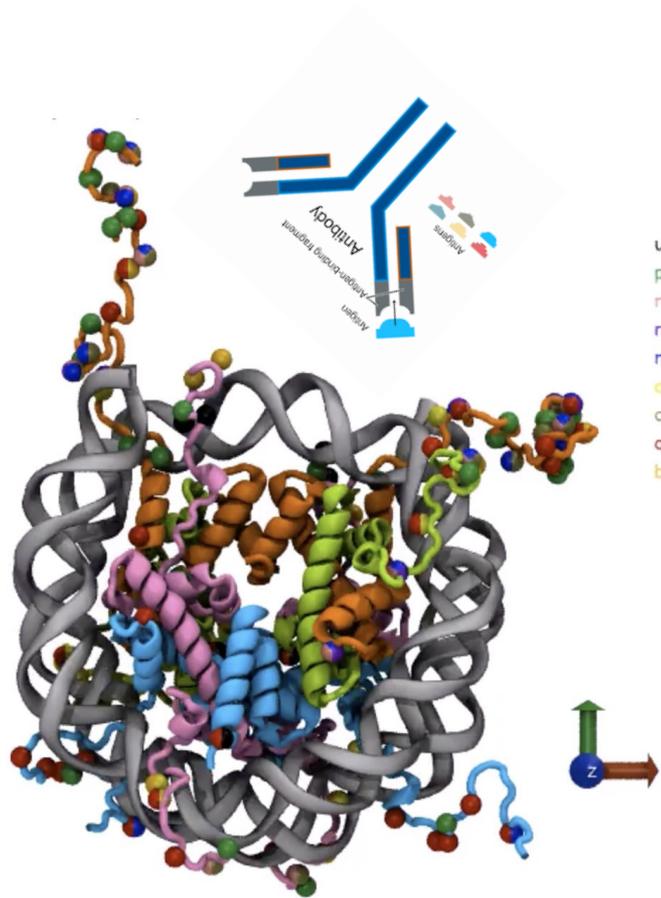
- Chromatin immunoprecipitation (ChIP)





ChIP-exo-seq





ub
ph
me2
me3
me1
ar
ci
ac
bio

H2A

H2AS1ph
H2AR3me2
H2AR3ci
H2AK5ac
H2AK9ac
H2AK9bio
H2AK13bio
H2AK13ar
H2AK119ub
H2AT120ph
H2AK121ub
H2AK125bio
H2AK127bio
H2AK129bio
H2AS137ph
H2AS139ph
H2AY142ph

H2B

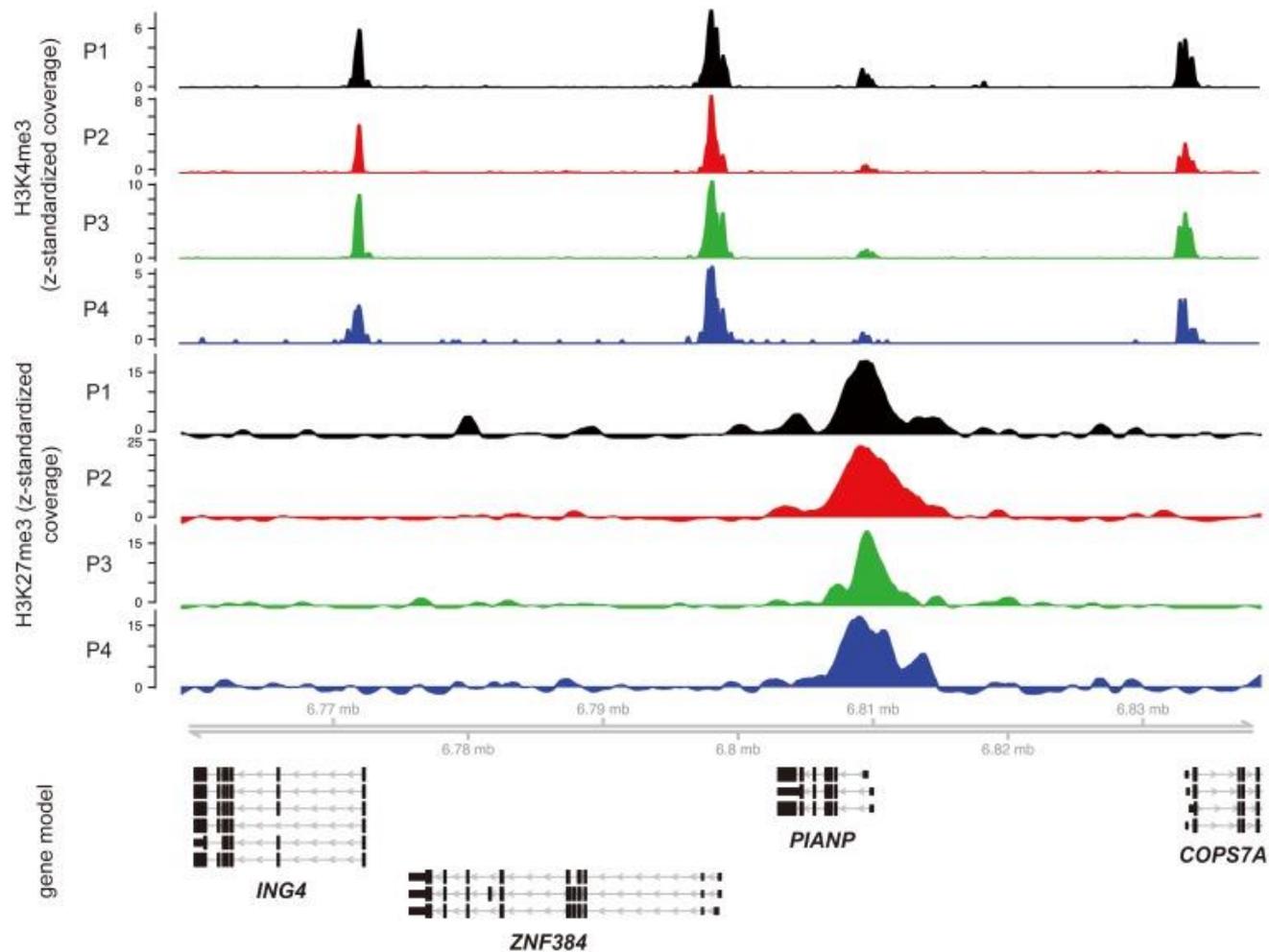
H2BK5ac
H2BK5me1
H2BK12ac
H2BS14ph
H2BK15ac
H2BK16ac
H2BK20ac
H2BK30ar
H2BK46ac
H2BK120ac
H2BK120ub

H3

H3R2me1
H3R2me2
H3R2ci
H3T3ph
H3K4ac
H3K4me1
H3K4me2
H3K4me3
H3S6ph
H3T6ph
H3R8ci
H3K9ac
H3K9me1
H3K9me2
H3K9me3
H3K9bio
H3S10ph
H3T11ph
H3K14ac
H3R17me1
H3R17me2
H3R17ci
H3K18ac
H3K18bio
H3K23ac
H3R26me1
H3R26ci
H3K27ac
H3K27me1
H3K27me2
H3K27me3
H3K27ar
H3S28ph
H3S31ph
H3K36ac
H3K36me3
H3K36me1
H3K36me2
H3K37ar
H3Y41ph
H3T45ph
H3K56ac
H3K79me1

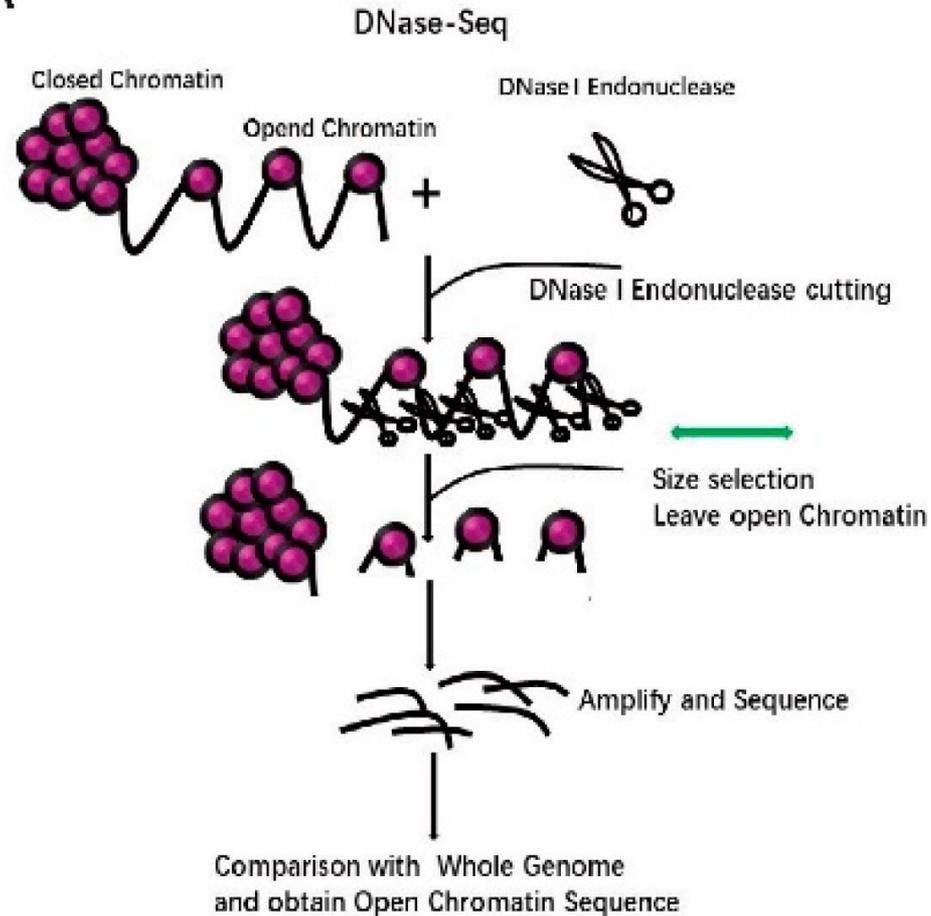
H4

H4S1ph
H4R3me1
H4R3ci
H4R3ci
H4K5ac
H4K8ac
H4K8bio
H4K12ac
H4K12bio
H4K16ac
H4K16ar
H4K20me1
H4K20me2
H4K20me3
H4K91ac
H4K91ub



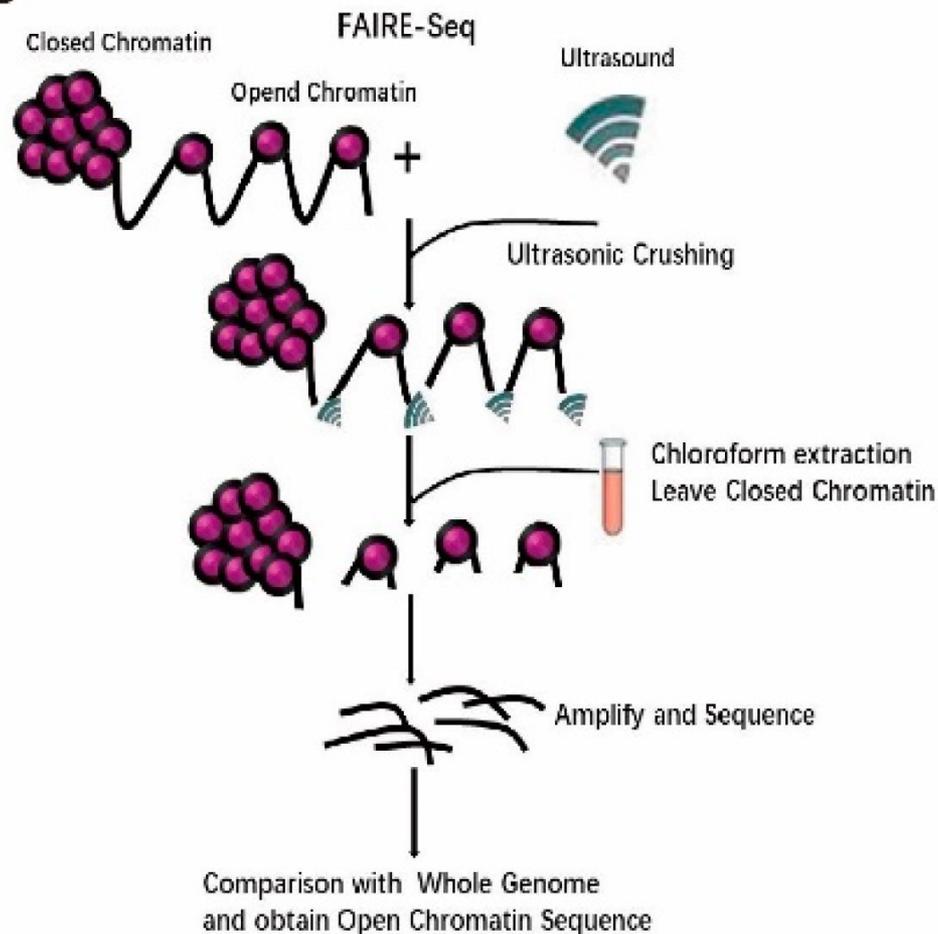
Определение доступности хроматина

A



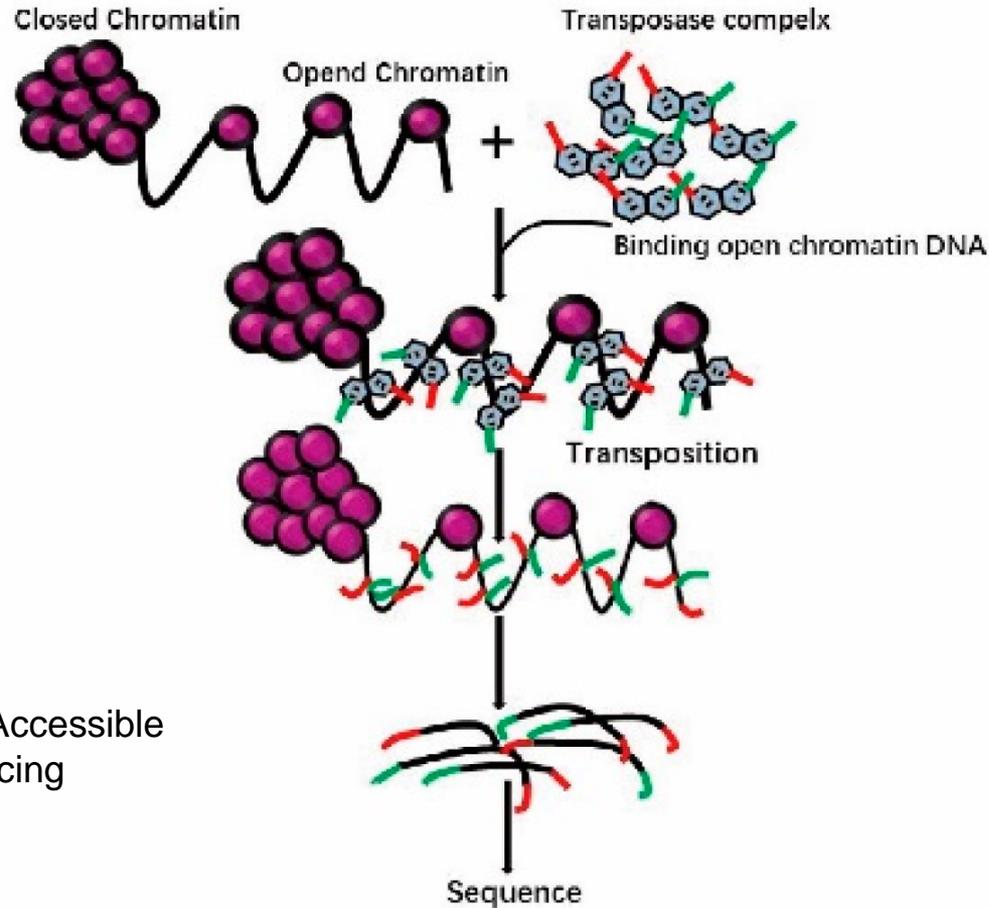
Определение доступности хроматина

B



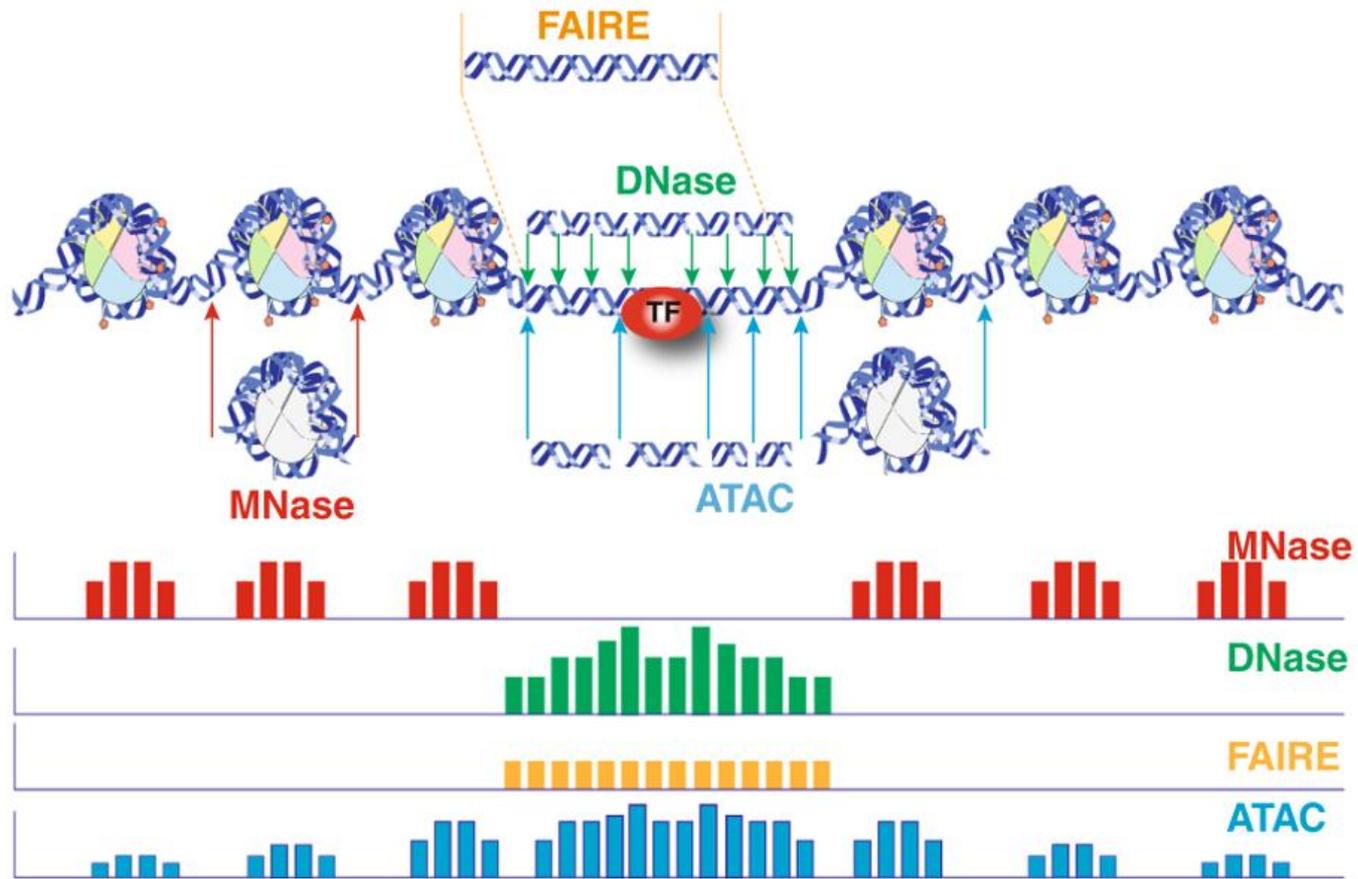
Определение доступности хроматина

ATAC-Seq

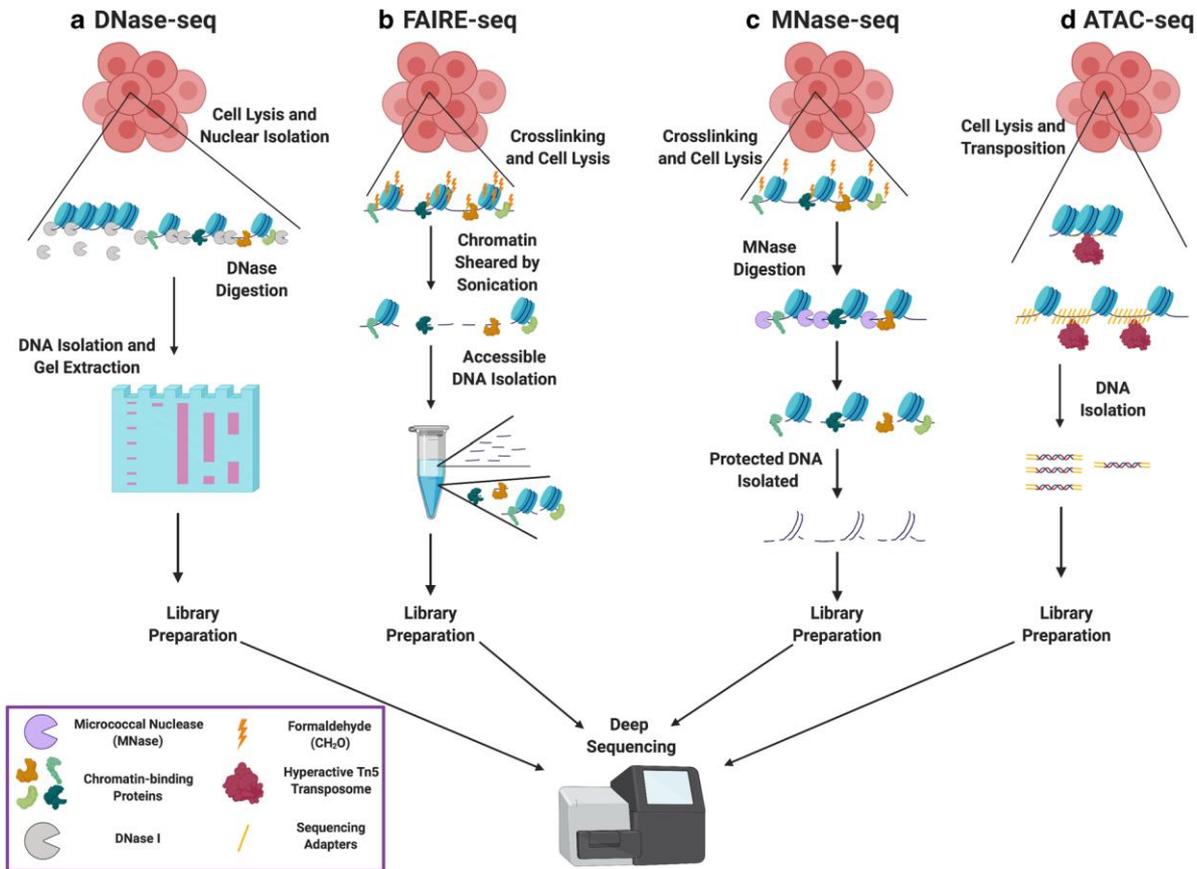


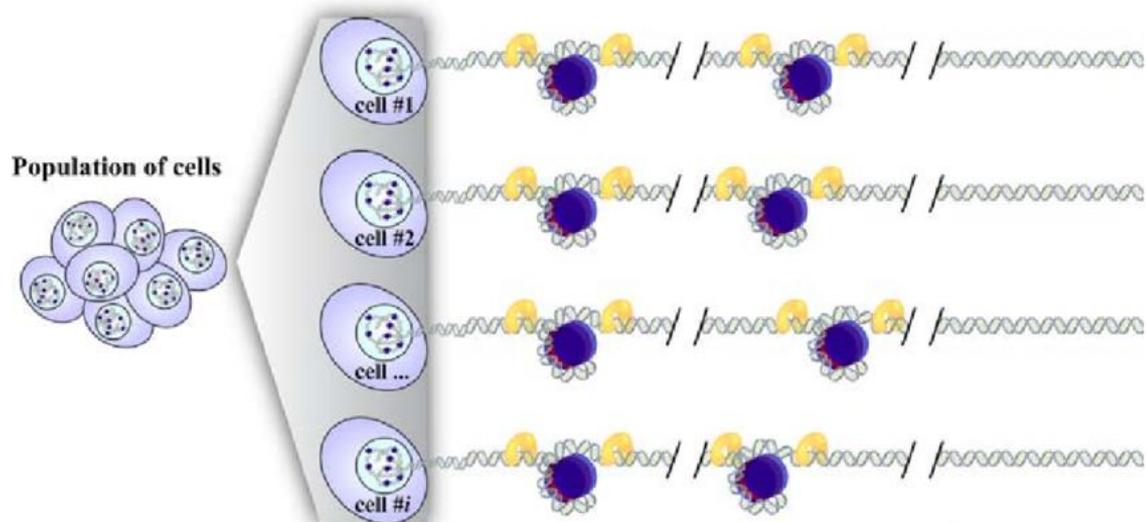
Assay for Transposase-Accessible Chromatin using sequencing

Определение доступности хроматина



Определение доступности хроматина

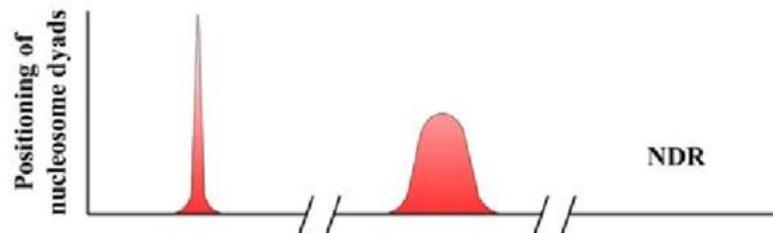




MNase-seq

nucleosomal DNA

Dyad



MNase-seq

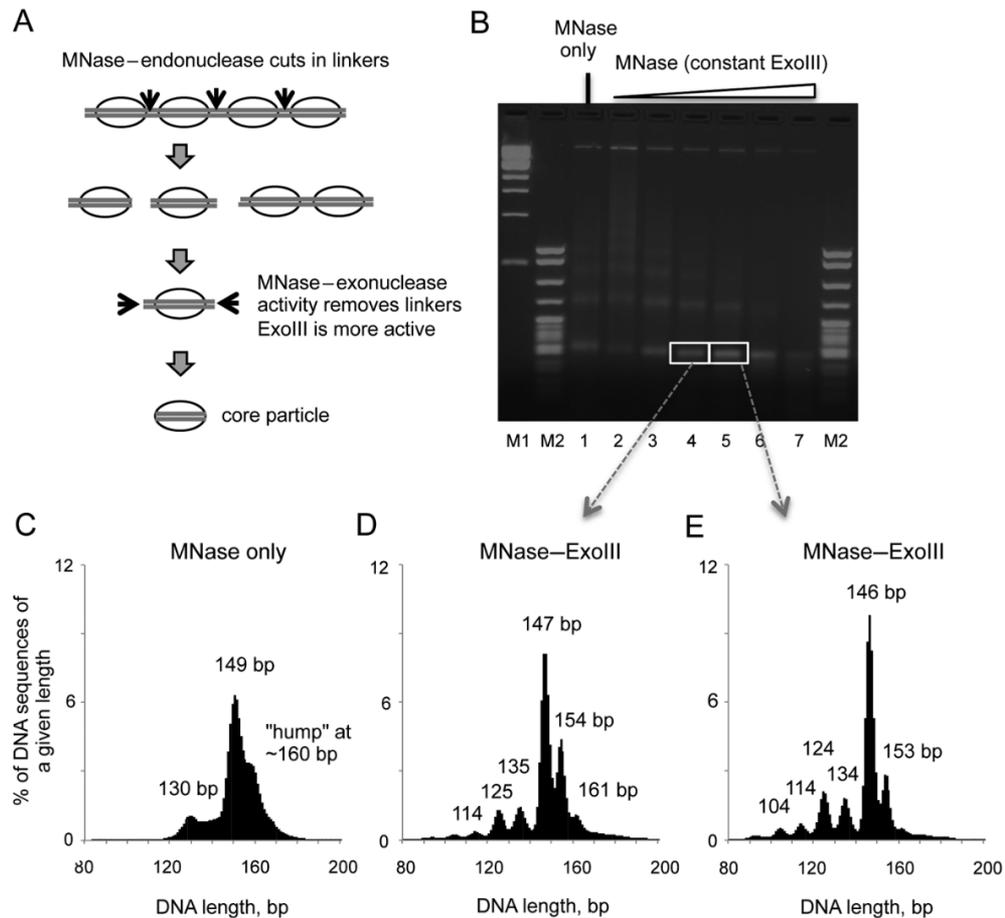
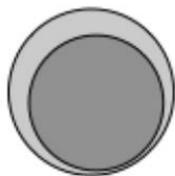
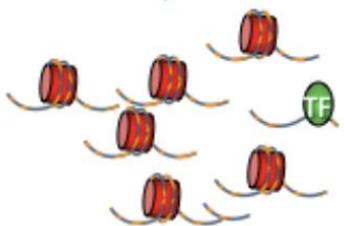


Figure 1. Simultaneous digestion of yeast chromatin with MNase and ExoIII results in a set of DNA fragments of discrete lengths, corresponding to the nucleosome core particle and its intermediates. (A) Digestion

A**ChIP**

↓ Crosslink and shatter



Specific antibody ↓

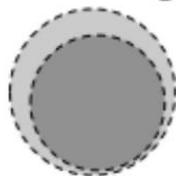
Precipitate ↓

Extract DNA ↓

Adapter ligation ↓

Amplify library ↓

Sequence

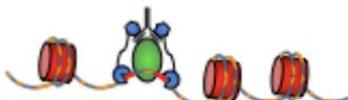
B**CUT&Tag**

permeabilize

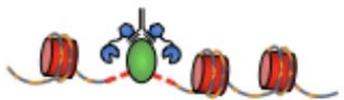
Specific antibody ↓



Protein A-Tn5 ↓



Mg⁺⁺ @ 37°C ↓



Extract DNA ↓

Amplify library ↓

Sequence

CUT&TAG

CUT&RUN

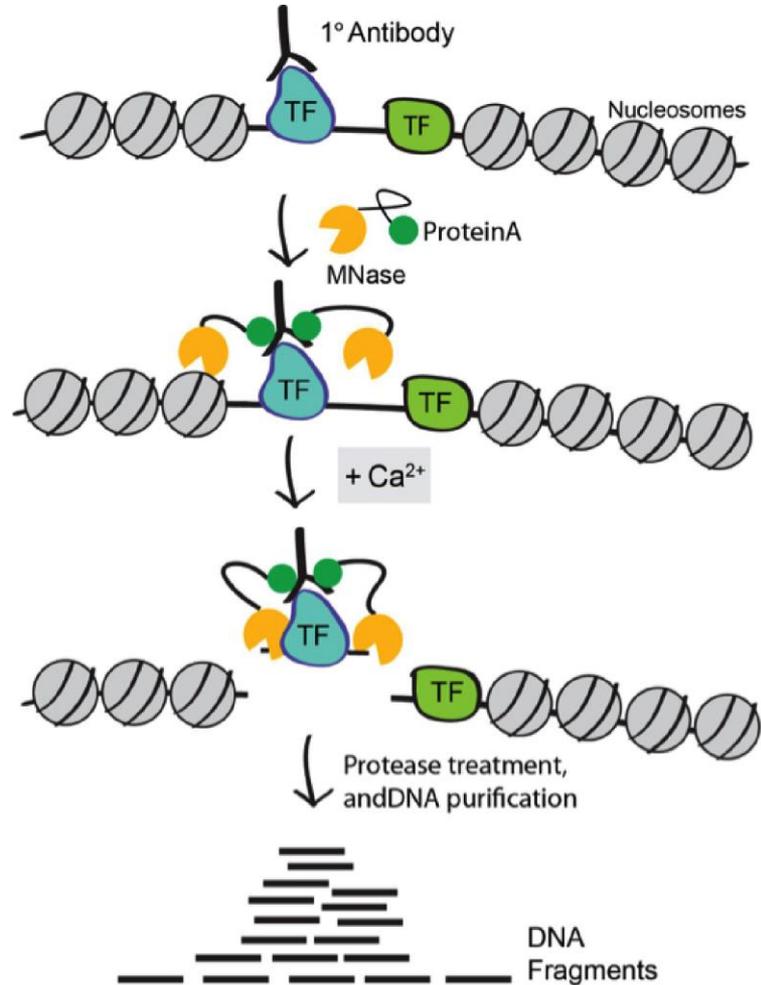
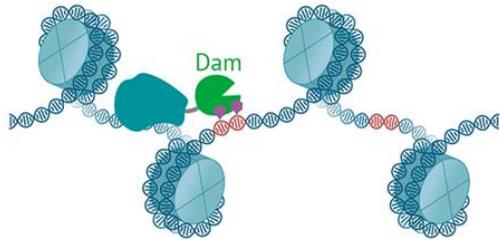


Figure 1. CUT&RUN schematic (see text for details).

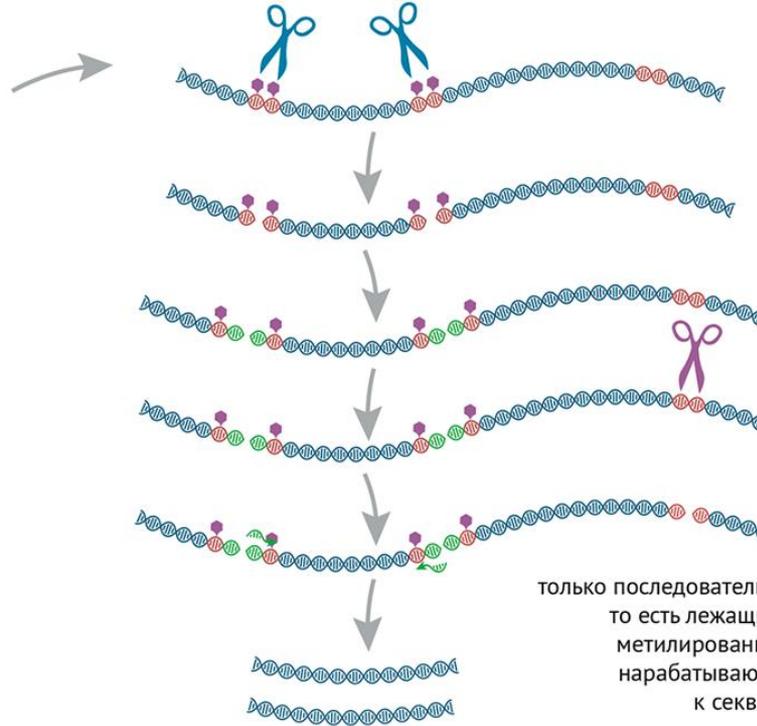
CUT&Tag vs. CUT&RUN vs. ChIP-Seq

	CUT&Tag	CUT&RUN	ChIP-Seq
Performed Under Native Conditions?	Yes	Yes	No
Chromatin Fragmentation Method	Tn5-based tagmentation	MNase digestion	Sonication
Cell Number Requirements	5,000-500,000 cells	500,000 cells	1-10 million cells
Sequencing Depth Required *	2 million reads **	8 million reads	20-50 million reads
Integrated Library Preparation?	Yes, uses tagmentation	No, separate library prep required	No, separate library prep required
Compatible Targets	Primarily histone modifications, some transcription factors and co-factors	Wide range of histone modifications, transcription factors, and co-factors	Wide range of histone modifications, transcription factors, and co-factors
Workflow Length	1-2 days	1-2 days	2-3 days

DamID – альтернатива ChIP-seq in vivo



ДНК метилтрансфераза **Dam** метилирует **GATC** последовательности на ДНК в тех местах хроматина, где находится гибридный белок, состоящий из интересующего нас **белка** и **Dam**



добавляют **рестриктазу** – “молекулярные ножницы” – она режет ДНК только по **метилированным GATC**-последовательностям

пришивают **адаптеры**

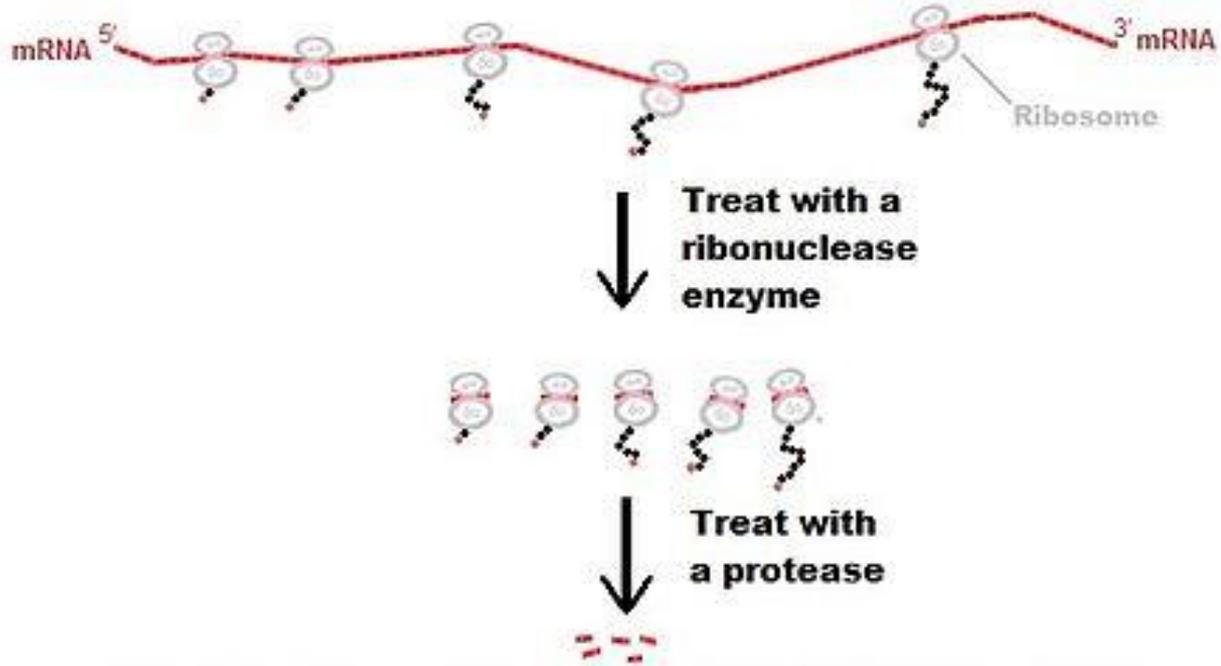
добавляют другую **рестриктазу**, которая режет ДНК только по **неметилированным GATC**-последовательностям

только последовательности с двумя адаптерами, то есть лежащие между соседними метилированными GATC-сайтами, нарабатываются ПЦР и готовятся к секвенированию

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6451315/>

<https://biomolecula.ru/articles/molekuly-i-epigenom#metody-izucheniia-dnk-i-khromatina>

Ribo-seq – определение мРНК, которые активно транслируются



Determine the sequence of the protected **RNA** fragments

Compare those sequences to the reference genome to determine **EXACTLY** where the ribosomes were.

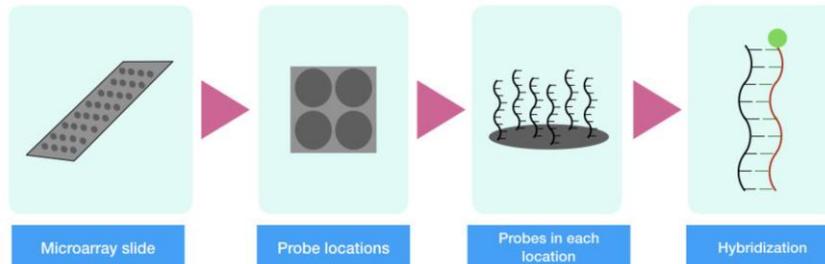
Методы ДНК микрочипов



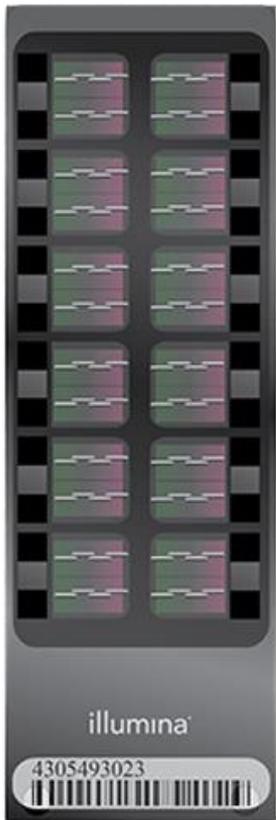
Андрей Дарьевич Мирзабеков



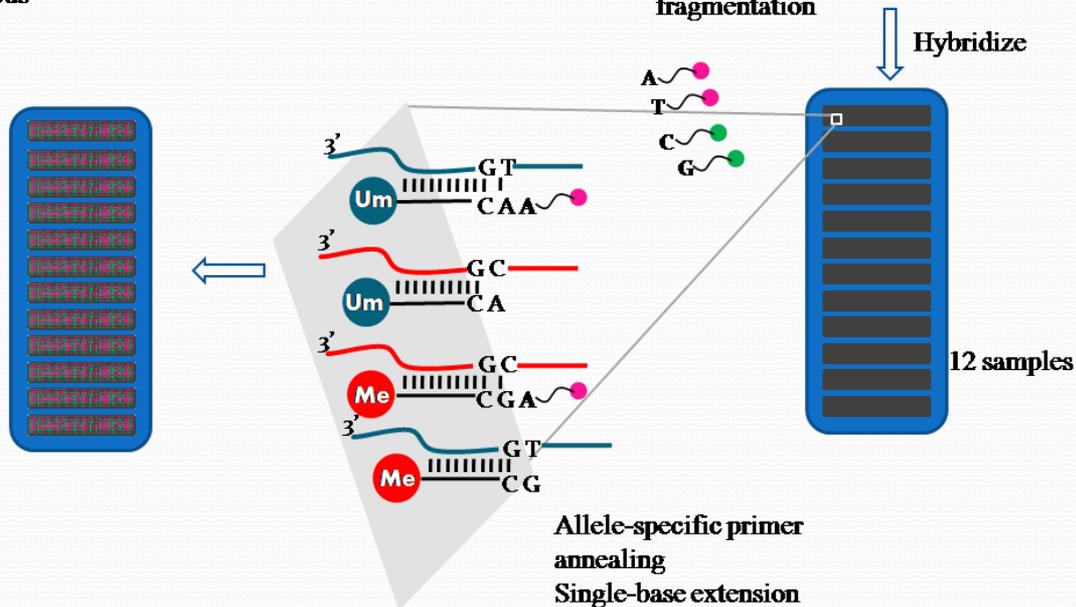
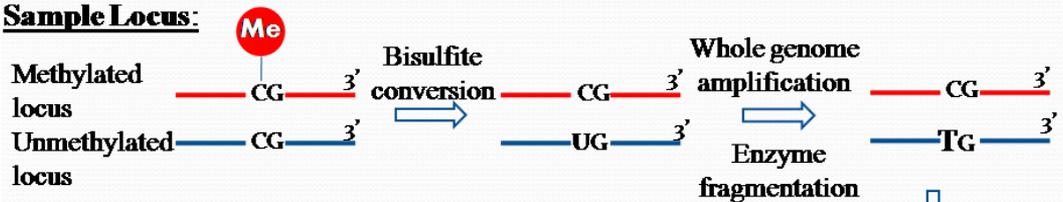
Дата рождения 19 октября 1937
Место рождения Баку, СССР
Дата смерти 13 июля 2003 (65 лет)



Illumina Infinium Methylation Assay



Sample Locus:



Infinium MethylationEPIC BeadChip ("EPIC") interrogates over 850,000 methylation sites across the human genome

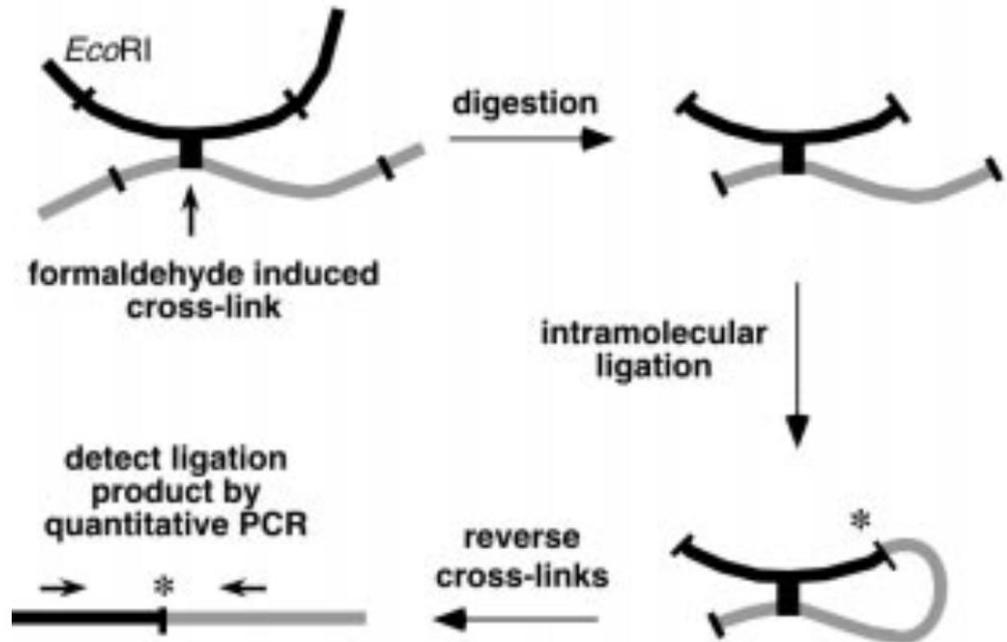
**3-C-методы – определение контактов ДНК в
геноме (3Д-укладка генома)**

CHROMOSOME CONFORMATION CAPTURE

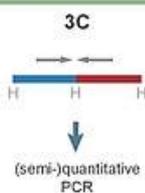
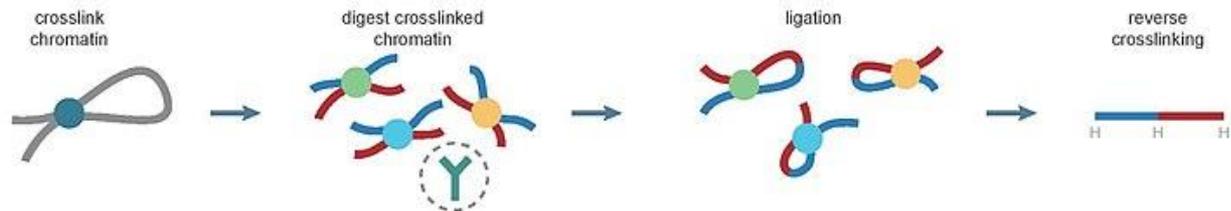
Capturing Chromosome Conformation

Job Dekker,^{1*} Karsten Rippe,² Martijn Dekker,³ Nancy Kleckner¹

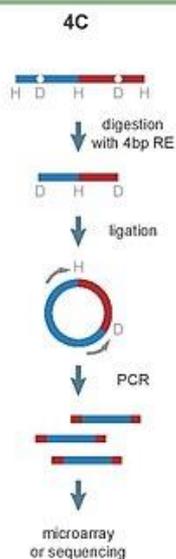
15 FEBRUARY 2002 VOL 295 SCIENCE



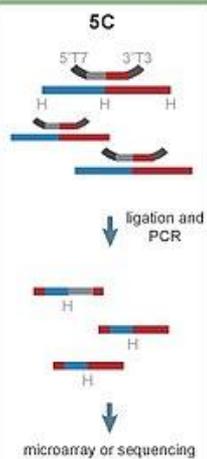
Chromosome Conformation Technologies



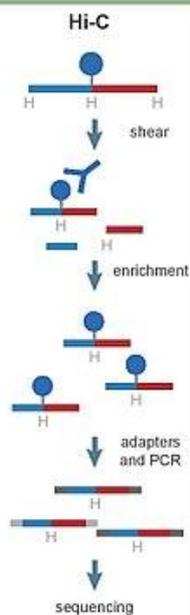
one vs one



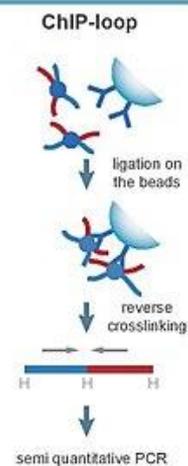
one vs all



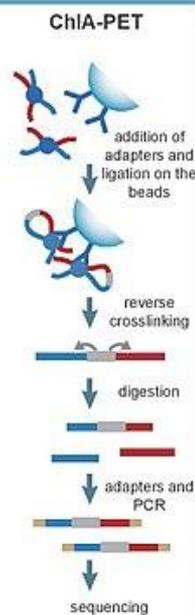
many vs many



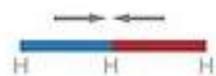
all vs all



one vs one



all vs all

3C

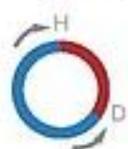
↓
(semi-)quantitative
PCR

4C

↓ digestion
with 4bp RE



↓ ligation



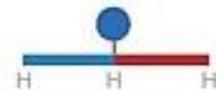
↓ PCR



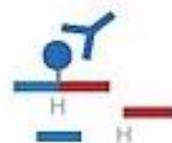
↓
microarray
or sequencing

one vs one

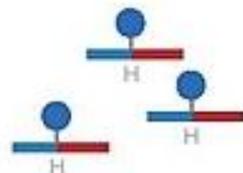
one vs all

Hi-C

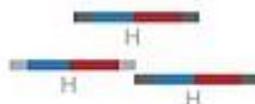
↓ shear



↓ enrichment



↓ adapters
and PCR



↓
sequencing

all vs all

ChIA-PET

↓ addition of
adapters and
ligation on the
beads



↓ reverse
crosslinking



↓ digestion



↓ adapters
and PCR

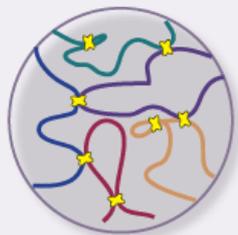


↓
sequencing

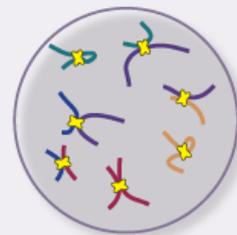
all vs all



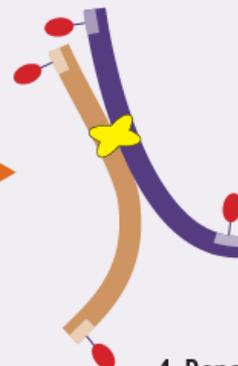
1. Start with cell or tissue samples



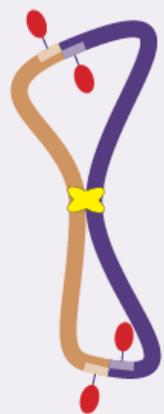
2. Crosslink chromatin



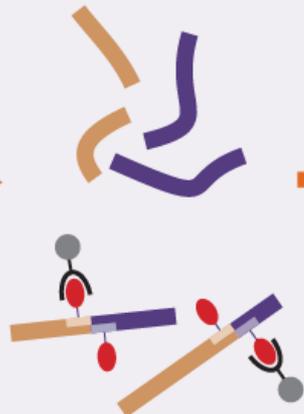
3. Fragment chromatin



4. Repair and biotinylate ends



5. Ligate fragmented chromatin



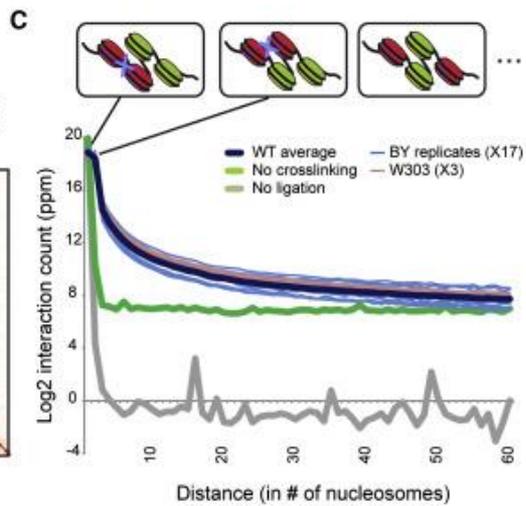
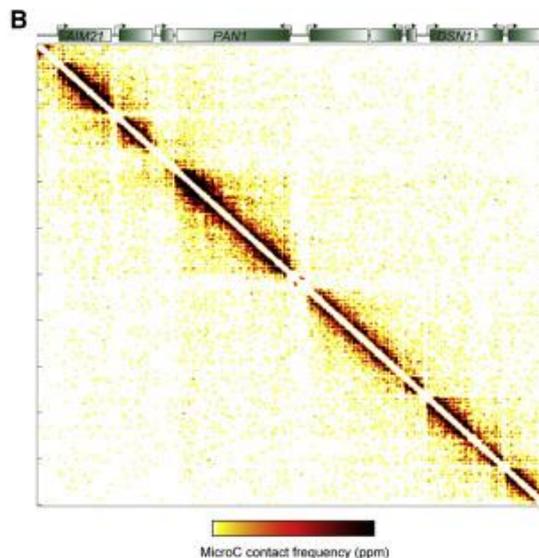
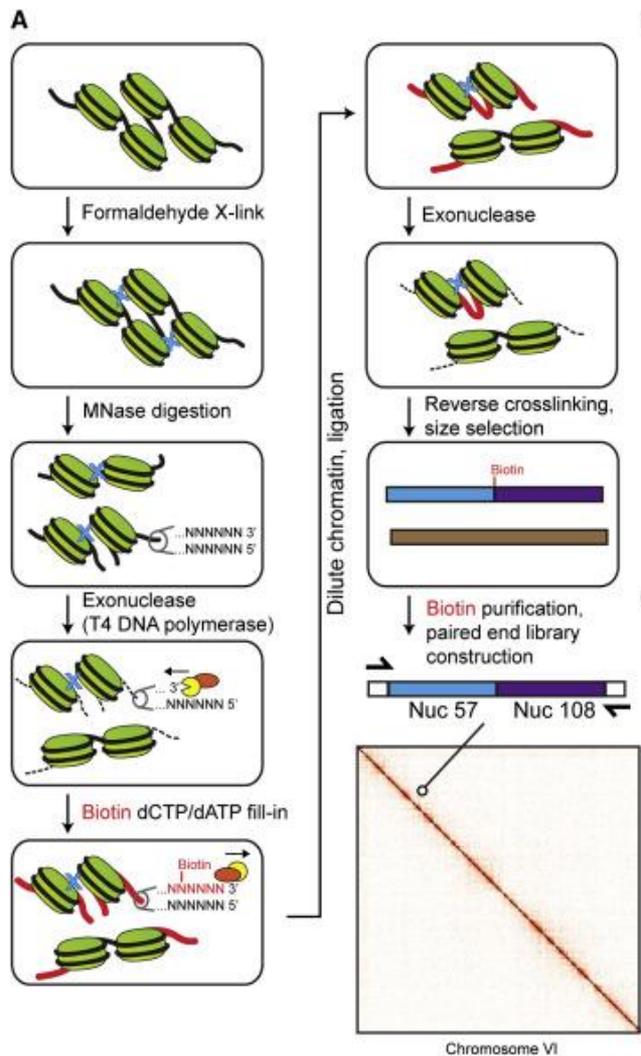
6. Shear DNA & pull down biotinylated DNA



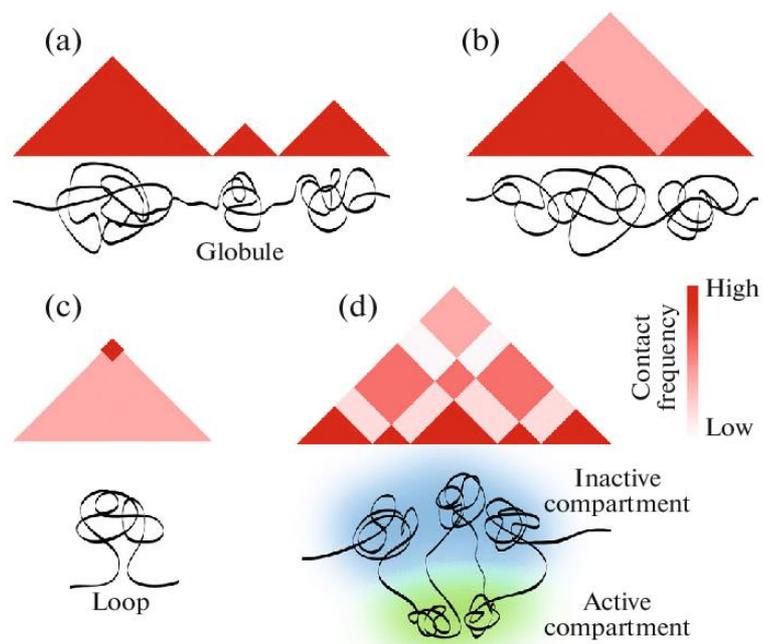
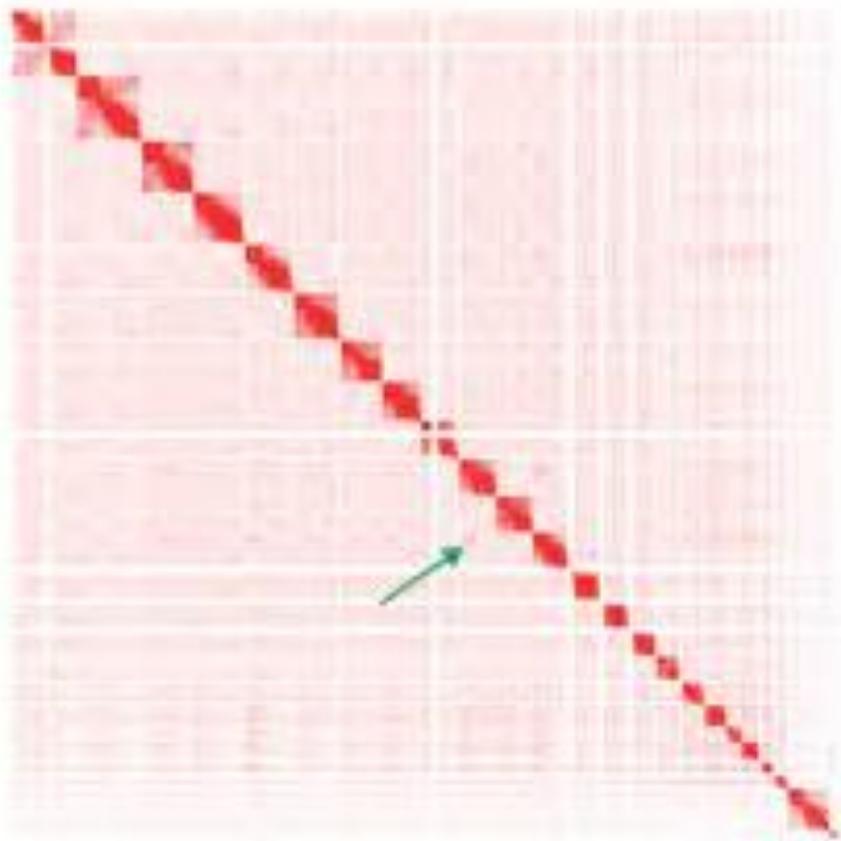
7. Paired-end Sequencing



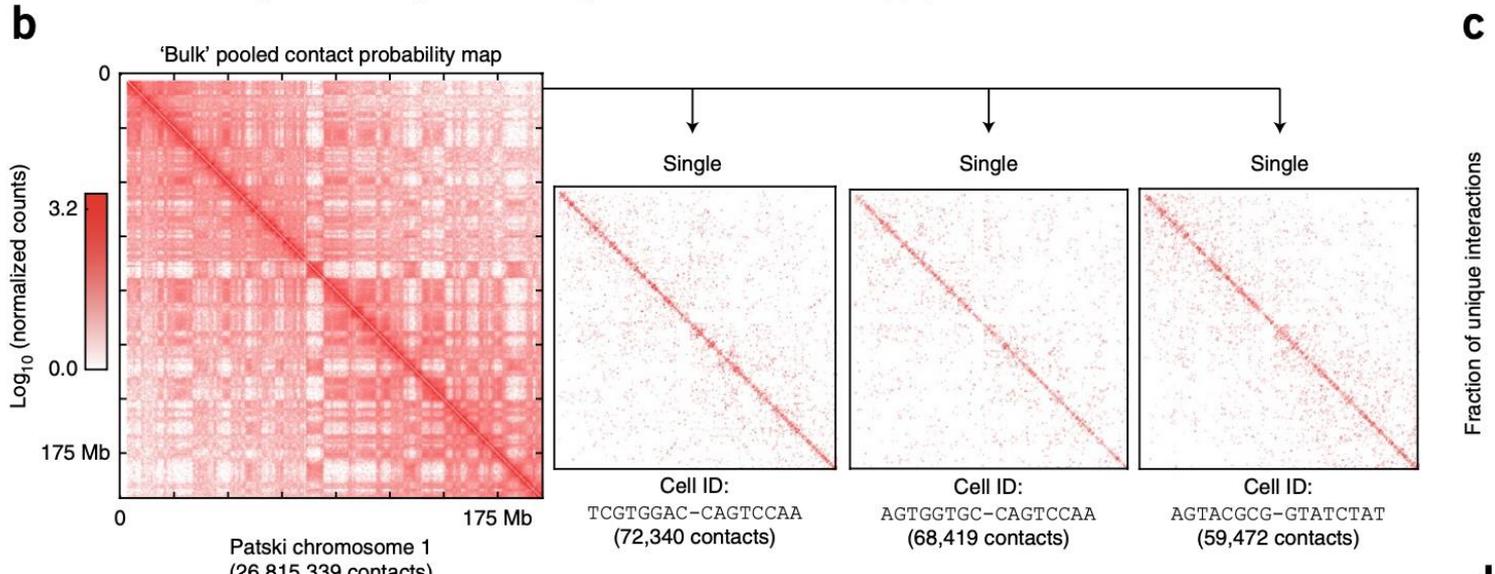
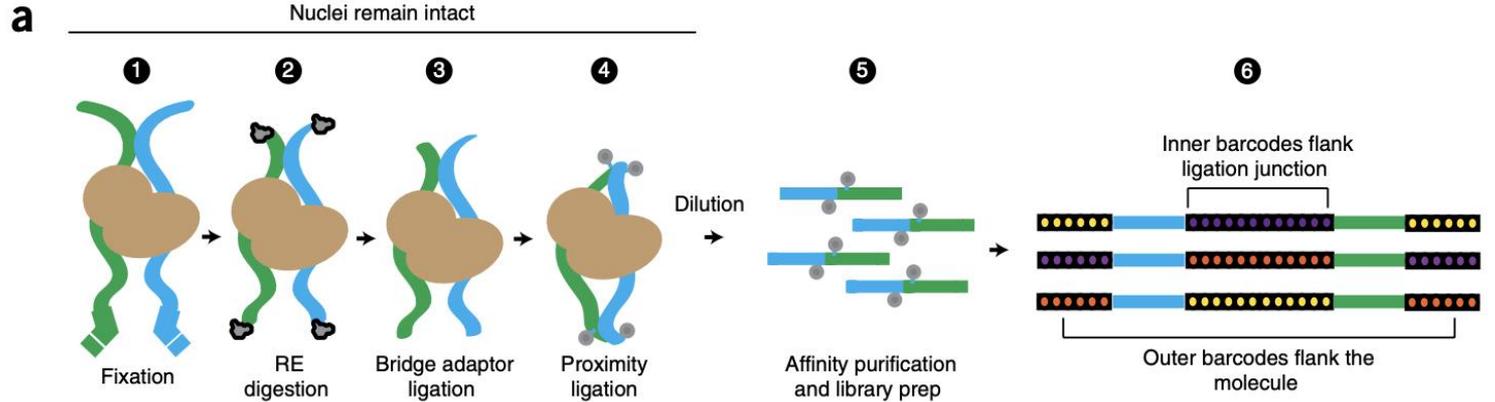
8. Bioinformatics



Micro-C

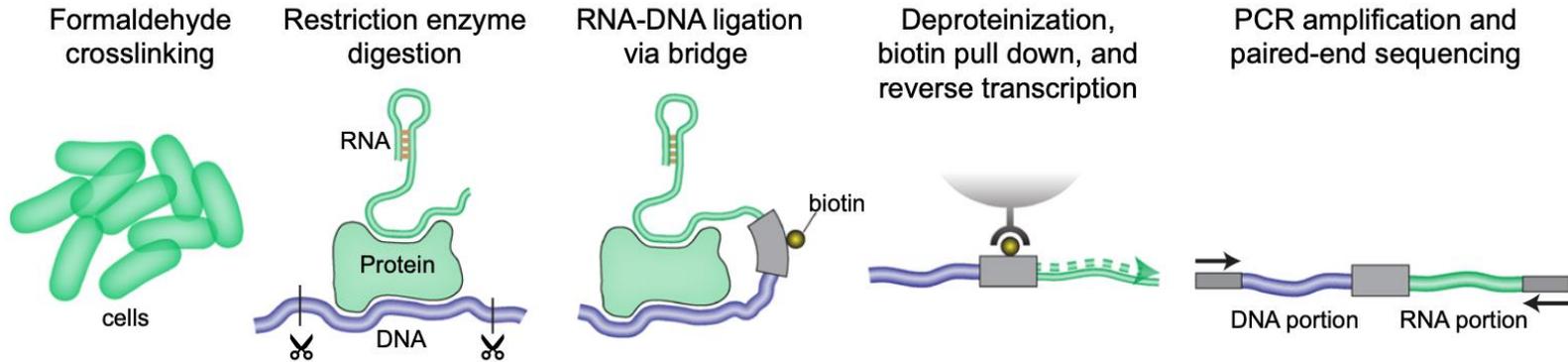


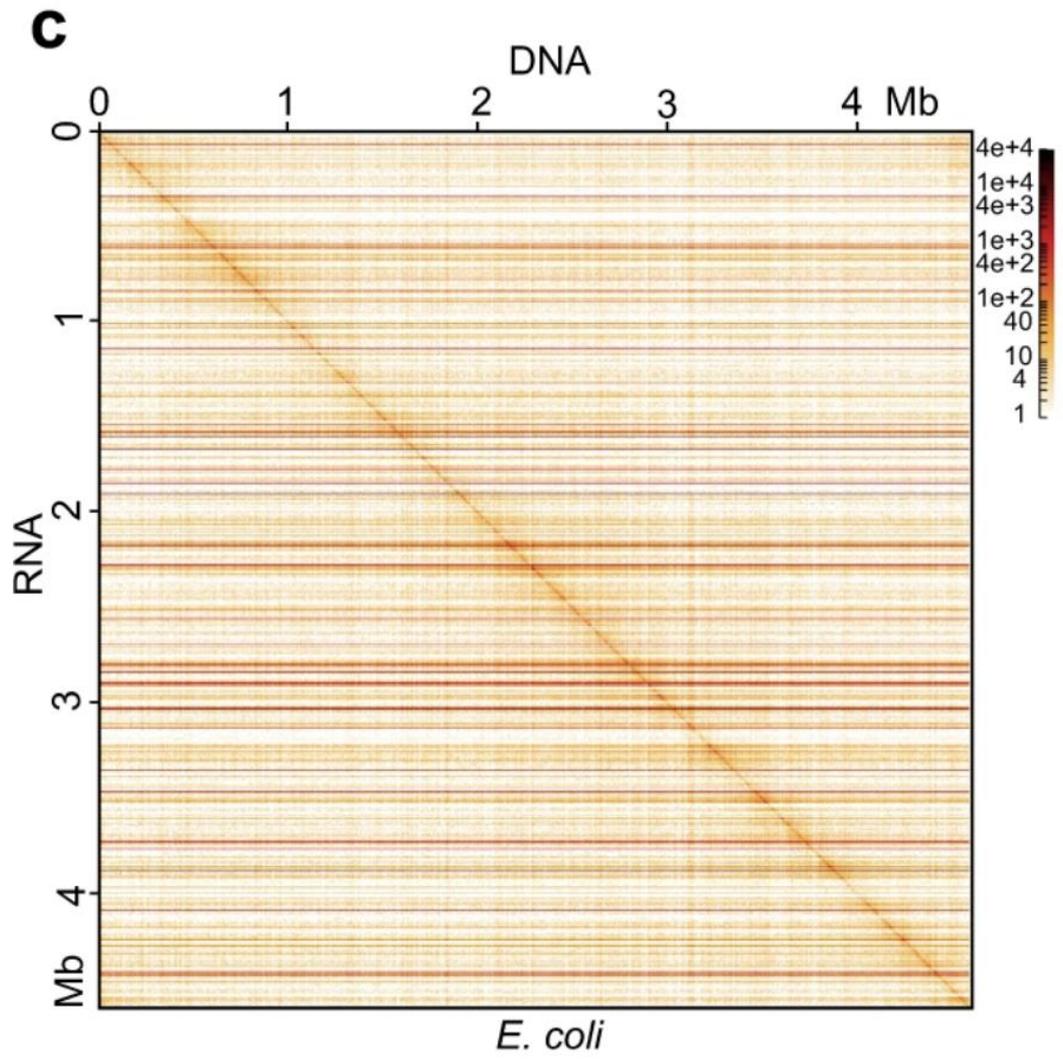
Single cell Hi-C



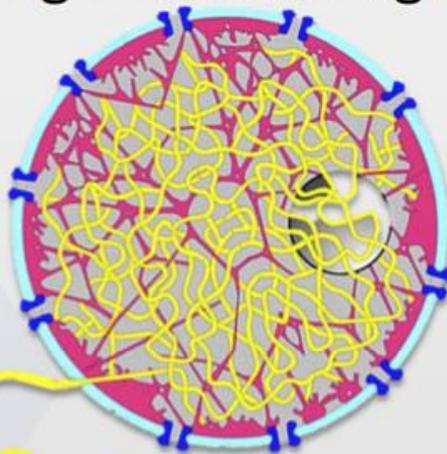
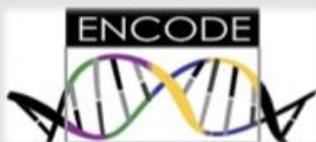
RNA-DNA interactomes, RedC method

a



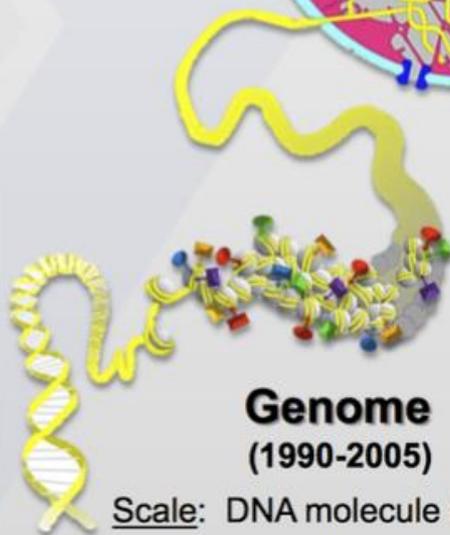


Finishing the Job: Understanding Genome Organization



3D Nucleome (2015-2022?)

Scale: cell nucleus &
chromatin domains



Epigenome (2005-2015)

Scale: nucleosome &
epigenetic marks

Genome (1990-2005)

Scale: DNA molecule &
sequence



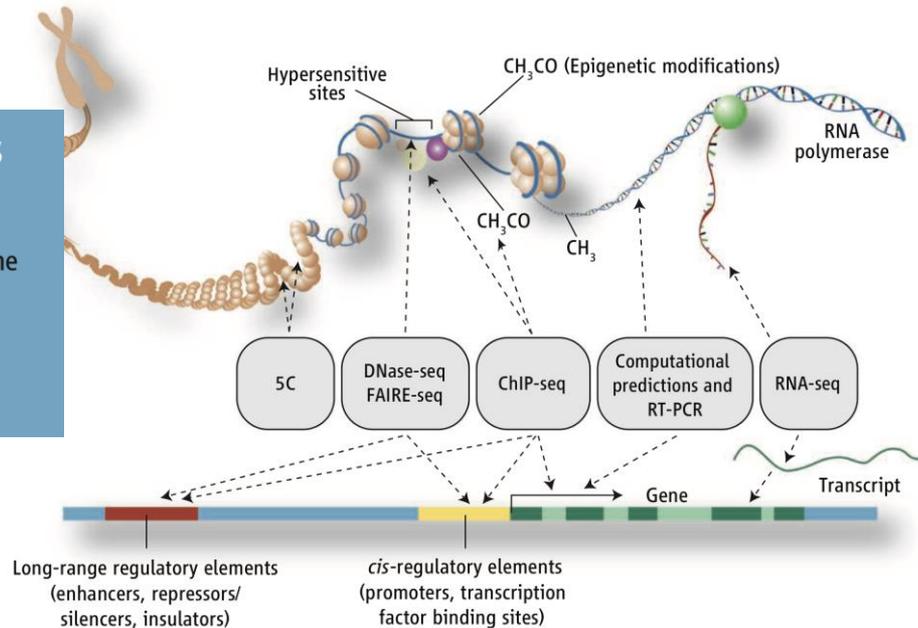
National Institutes of Health
Office of Strategic Coordination - The Common Fund

ENCODE Project Writes Eulogy For Junk DNA

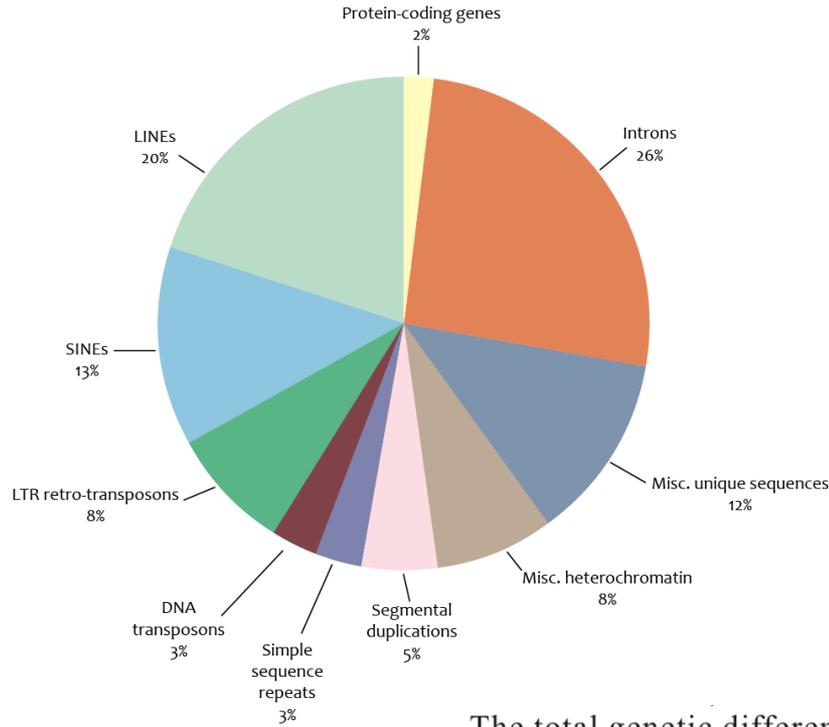
SCIENCE VOL 337 7 SEPTEMBER 2012

ENCODE By the Numbers

- 147 cell types studied
- 80% functional portion of human genome
- 20,687 protein-coding genes
- 18,400 RNA genes
- 1640 data sets



Структура генома человека и вариации



4 to 5 million SNPs in a person's genome

The total genetic difference between humans and chimps, in terms of number of bases, sums to about 4% of the genome. That



2 SEPTEMBER 2005 VOL 309 SCIENCE

99% identity of the aligned sequence
96% identity between whole genomes

On the Immortality of Television Sets: “Function” in the Human Genome According to the Evolution-Free Gospel of ENCODE

Dan Graur , Yichen Zheng, Nicholas Price, Ricardo B.R. Azevedo, Rebecca A. Zufall, Eran Elhaik [Author Notes](#)

Genome Biology and Evolution, Volume 5, Issue 3, March 2013, Pages 578–590,
<https://doi.org/10.1093/gbe/evt028>

Published: 20 February 2013 **Article history** ▼

Whatever your proposed functions are, ask yourself this question: Why does an onion need a genome that is about five times larger than ours?”

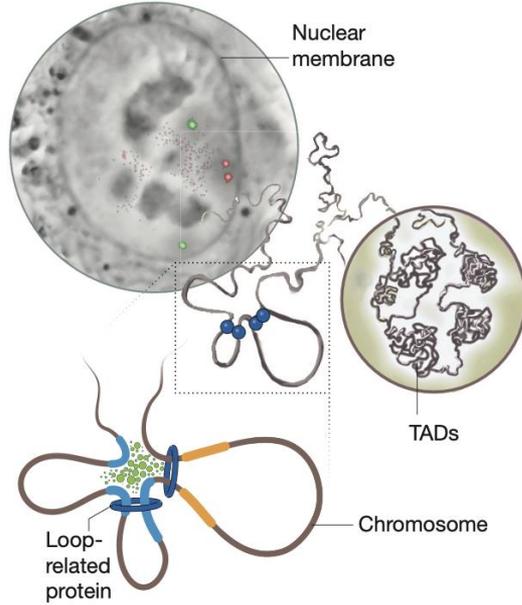
—T. Ryan Gregory (personal communication)

Actually, the ENCODE authors could have chosen any of a number of arbitrary percentages as “functional,” and . . . they did! In their scientific publications, ENCODE promoted the idea that 80% of the human genome was functional. The scientific commentators followed, and proclaimed that at least 80% of the genome is “active and needed” (Kolata 2012). Subsequently, one of the lead authors of ENCODE admitted that the press conference misled people by claiming that 80% of our genome was “essential and useful.” He put that number at 40% (Gregory 2012), although another lead author reduced the fraction of the genome that is devoted to function to merely 20% (Hall 2012). Interestingly, even when a lead author of ENCODE reduced the functional genomic fraction to 20%, he continued to insist that the term “junk DNA” needs “to be totally expunged from the lexicon,” inventing a new arithmetic according to which $20\% > 80\%$. In its synopsis of the year 2012, the journal *Nature* adopted the more modest estimate, and summarized the findings of ENCODE by stating that “at least 20% of the genome can influence gene expression” (Van Noorden 2012). *Science* stuck to its maximalist guns, and its summary of 2012 repeated the claim that the “functional portion” of the human genome equals 80% (Anonymous 2012). Unfortu-

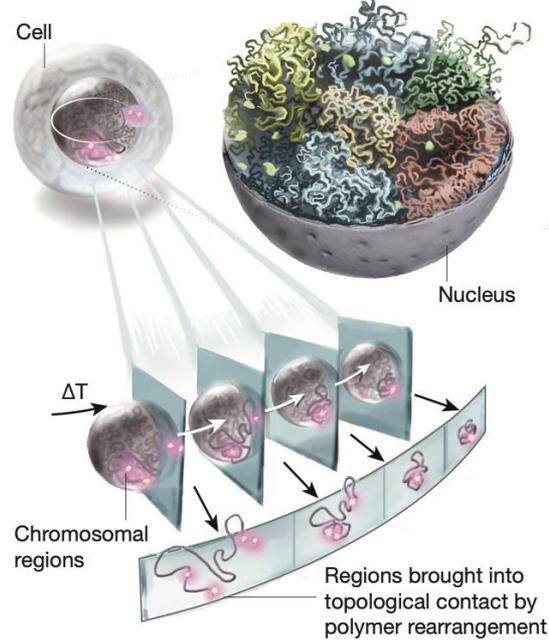
- A recent slew of ENCYclopedia Of DNA Elements (ENCODE) Consortium publications, article signed by all Consortium members, put forward the idea that more than 80% genome is functional. This claim flies in the face of current estimates according to of the genome that is evolutionarily conserved through purifying selection is less according to the ENCODE Consortium, a biological function can be maintained selection, which implies that at least $80 - 10 = 70\%$ of the genome is perfectly deleterious mutations, either because no mutation can ever occur in these because no mutation in these regions can ever be deleterious. This absurd conclusion through various means, chiefly by employing the seldom used “causal role” function and then applying it inconsistently to different biochemical properties, by logical fallacy known as “affirming the consequent,” by failing to appreciate the between “junk DNA” and “garbage DNA,” by using analytical methods that yield biased errors and inflate estimates of functionality, by favoring statistical sensitivity over specificity, and by emphasizing statistical significance rather than the magnitude of the effect. Here, we detail the many logical and methodological transgressions involved in assigning functionality to almost every nucleotide in the human genome. The ENCODE results were predicted by one of its authors to necessitate the rewriting of textbooks. We agree, many textbooks dealing with marketing, mass-media hype, and public relations may well have to be rewritten.

- Недавнее множество публикаций Консорциума ENCCyclopedia Of DNA Elements (ENCODE), особенно статья, подписанная всеми членами Консорциума, выдвинула идею о том, что более 80% генома человека является функциональным. Это заявление идет вразрез с современными оценками, согласно которым доля генома, эволюционно законсервированного посредством очищающего отбора, составляет менее 10%. Таким образом, согласно консорциуму ENCODE, биологическая функция может поддерживаться неопределенно долгое время без селекции, что означает, что по крайней мере $80 - 10 = 70\%$ генома совершенно неуязвимы для вредоносных мутаций либо потому, что никакая мутация никогда не может произойти в этих «функциональных» или потому, что никакая мутация в этих регионах никогда не может быть вредной. Этот абсурдный вывод был сделан различными способами, главным образом путем использования редко используемого определения «каузальной роли» биологической функции, а затем непоследовательного применения его к различным биохимическим свойствам, путем совершения логической ошибки, известной как «подтверждение следствия», из-за неспособности оценить ключевое различие между «junk DNA» и «garbage DNA», используя аналитические методы, которые дают предвзятые ошибки и завышают оценки функциональности, отдавая предпочтение статистической чувствительности, а не специфичности, и подчеркивая статистическую значимость, а не величину эффекта. Здесь мы подробно описываем множество логических и методологических ошибок, связанных с определением функциональности почти каждого нуклеотида в геноме человека. Один из авторов предсказал, что результаты ENCODE потребуют переписывания учебников. Мы согласны с тем, что многие учебники по маркетингу, шумихе в СМИ и связям с общественностью, возможно, придется переписать.

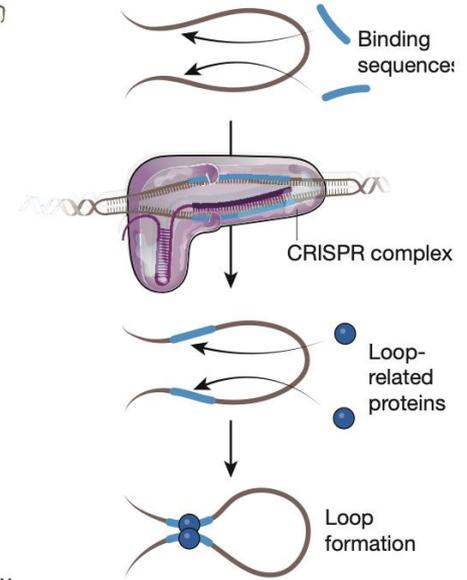
a Mapping – molecular genomic mapping of contacts, imaging of contacts and dynamics



b Model building – by constraint modelling and polymer simulation



c Functional validation – by genetic and biophysical perturbation experiments

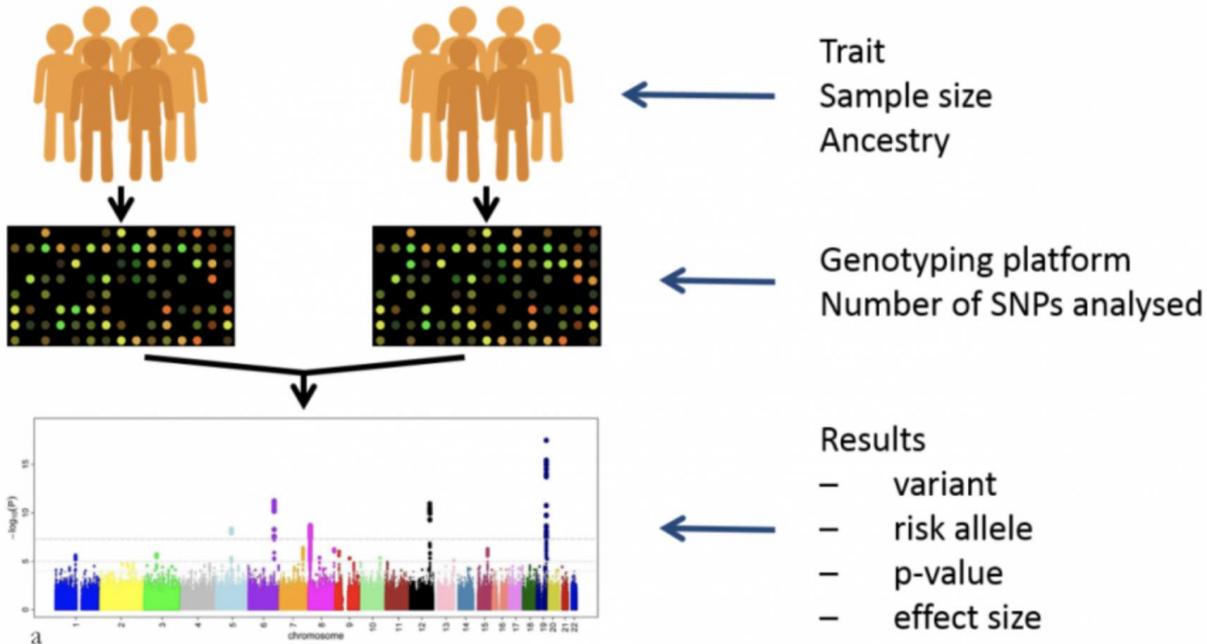


4D Nucleome

<https://www.4dnucleome.org>

GWAS

Genome-wide association study



Manhattan plot



dpGAP

<https://www.ncbi.nlm.nih.gov/gap>

CIDR: Collaborative Study on the Genetics of Alcoholism Case Control Study

dbGaP Study Accession: phs000125.v1.p1

[Request Access](#)

[Show BioProject list](#)

[Study](#) [Variables](#) [Documents](#) [Analyses](#) [Datasets](#) [Molecular Data](#)

Analysis Name and Accession

Name: GWAS for alcohol dependence in European-Americans
Accession: pha002892.1

[View association results in Genome Browser](#)

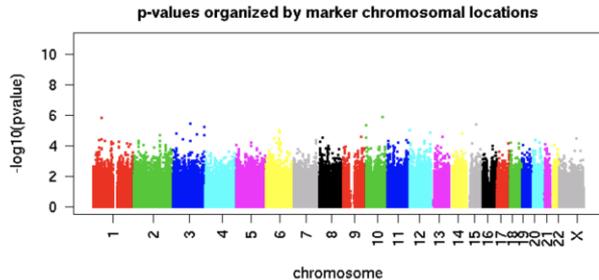
Analysis Description

To identify common variants underlying alcohol-dependence, probands were ascertained through alcohol treatment programs and evaluated and their relatives were administered a validated poly-diagnostic instrument, the Semi- Structured Assessment for the Genetics of Alcohol performed by the Center for Inherited Disease Research (CIDR). DNA sources included blood (n = 1453) and lymphoblastoid cell lines

Analysis Methods

Sample QC filters consisted of 98% genotyping completeness, unrelatedness of subjects (n = 9). A principal component-based analysis of either the European or African-American groups and were excluded. EA and AA SNP QC filters (applied separately in each group) were performed in PLINK adjusting for sex.

Analysis Plots



dbGaP Genome Browser: pha002892

Display Option: [-log₁₀ P-value](#) [Filtering](#) [Zoom](#)

[-log₁₀ P-value](#)

[N/A](#) [-4](#) [4](#) [-8](#) [8](#) [-16](#) [16](#) [-32](#) [32](#) [-64](#) [64](#) [-128](#) [128](#) [-256](#) [256](#) [-512](#) [512](#) [-1024](#) [1024](#)

[Search by context](#) [Agenda](#) [Feedback](#)

ANALYSIS: GWAS for alcohol dependence in European-Americans
METHOD: categorical analysis
STUDY: CIDR: Collaborative Study on the Genetics of Alcoholism (COGA) (pha000125)

[Download the displayed data](#)

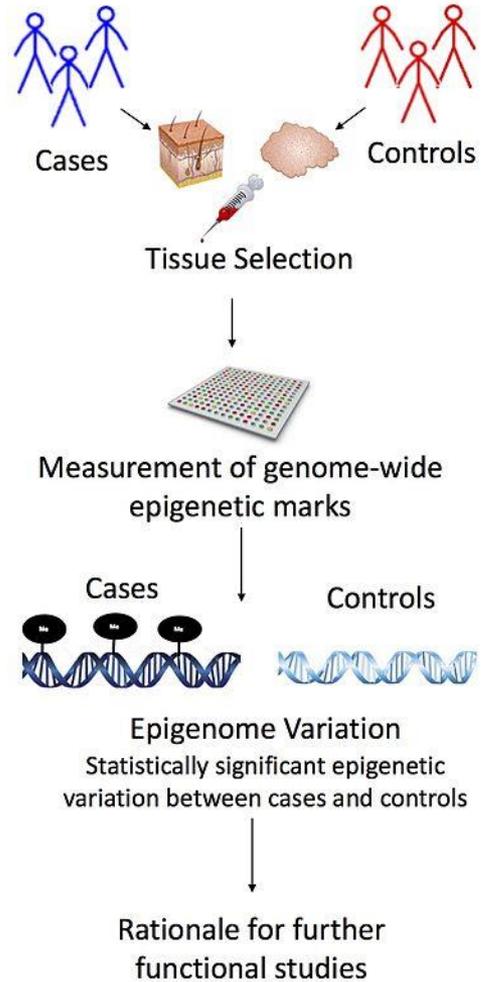
For full data set, please apply through [Authorized Access](#)

SNP bins on chromosomes:
 Chromosomes on this page are linearly divided into SNP bins in length of 2 million bases, and the SNP bin color corresponds to the P-value of top hit marker(s) when applicable.

Mouse-over of SNP bins:
 Mouse-over of a SNP bin shows the number of top SNP hits and of total SNP hits along with the bin genome co-ordinates.

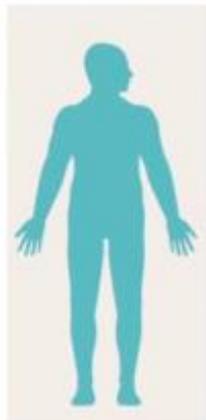
Click a SNP bin:
 Click a SNP bin to open graphic Gap Browser for details of genotype and phenotype for SNP markers as well as references/utility tools.

Epigenome-wide association study (EWAS)

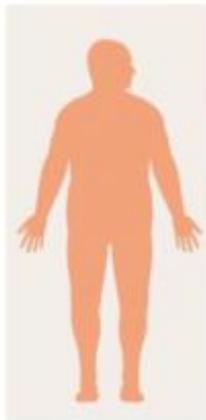


Meta-EWAS of incident type 2 diabetes

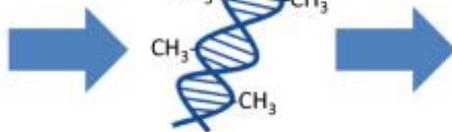
7–10 years prior



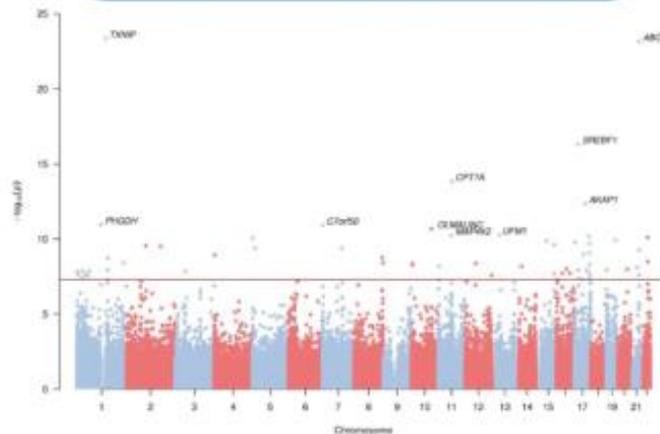
Type 2 diabetes

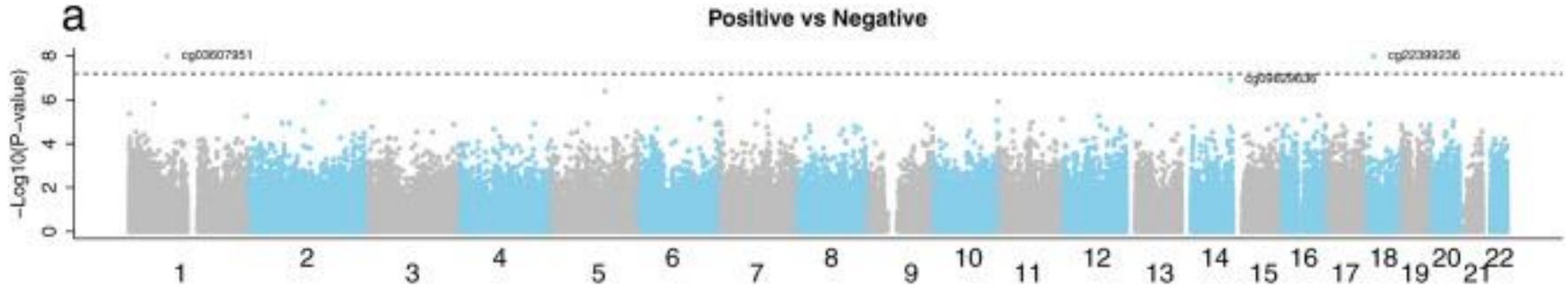


Blood



DNA methylation





[Sci Rep.](#) 2022; 12: 11478.

PMCID: PMC9261254

Published online 2022 Jul 7. doi: [10.1038/s41598-022-15467-1](https://doi.org/10.1038/s41598-022-15467-1)

PMID: [35798818](https://pubmed.ncbi.nlm.nih.gov/35798818/)

EWAS of post-COVID-19 patients shows methylation differences in the immune-response associated gene, *IFI44L*, three months after COVID-19 infection

[Yunsung Lee](#),^{#1} [Espen Riskedal](#),^{#2} [Karl Trygve Kalleberg](#),² [Mette Istre](#),³ [Andreas Lind](#),⁴ [Fridtjof Lund-Johansen](#),⁵ [Olaug Reiakvam](#),³ [Arne V. L. Søråas](#),³ [Jennifer R. Harris](#),¹ [John Arne Dahl](#),³ [Cathrine L. Hadley](#),² and [Astanand Jugessur](#)^{1,6}

Спасибо за внимание!